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November, 2021 Volume 3, Issue 2

A rare and endemic species expanded to Northern Anatolia: Grupocoris syriacus Reuter, 1896 (Hem., Het., Miridae)

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ABSTRACT: In this paper, new record distributional data on a rare Heteroptera (Hemiptera) species, Grypocoris syriacus Reuter, 1896 from northern Anatolia (Gerede district, Bolu province) are given.

KEYWORDS: Grypocoris syriacus Reuter 1896, Hemiptera, rare endemic species, Turkey.

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INTRODUCTION

New studies have been added over the years regarding the known number and On the other hand, the species found distribution of Heteroptera in Turkey. In naturally in only one geographical area is this way, while the number of newly recorded species from the fauna of Turkey increases, the distribution limits of the existing species in the fauna also expand.

it lives and found in a geographically narrow naturally area.

an endemic species.

In this study, the literatures on the taxonomy and distribution of Grypocoris syriacus Reuter 1896 was reviewed (Aukema & Rieger, 1999; Carvalho, 1959; In general, a species is considered rare if Dursun & Fent, 2017; Hoberlandt, 1956; Kerzhner, 1997; Kerzhner & Matocq, 1994; MATERIAL AND METHODS Önder, et al., 2006; Reuter, 1896a,b; Stichel, 1957-1962; Wagner, 1966; Wagner, 1970/71).

Reuter 1896 has been limited to a local area in 2017 (Table 1). geographical area only (Eastern Medi-terranean province of Anatolia). In which case this species is "locally endemic". The G.syriacus Reuter, 1896 is also both rare and anatolian endemic species.

endemic species G. syriacus Reuter, 1896, using identification keys by Stichel (1957 expanded its geographical distribution, -1962), and Wagner (1970/71, 1974) by and it was determined that this species second author. expanded to Northern Anatolia.

This study was conducted in Gerede surrounding of Bolu province, a total of 6 individual specimens of the species were Until now, the distribution of G.suriacus collected from 2 locations in the study

The specimens were collected by sweep net on the herbaceous vegetation by first author. And killed in 70% alcohol jars and were prepared based on technical and standards of data collection of the According to this study, the rare and zoology museum. These were determined

Table 1. Sampling localities of specimens G.syriacus Reuter, 1896 from Gerede/Bolu.

Locality No	Sampling localities	Coordinates	Altitude
Loc.1-S10	Avcılar mahallesi	40°48'16"N 32°28'28"E	1261m
Loc.2-S12	Kayısopran Köyü	40°48'18"N 32°27'13"E	1344m

RESULTS

Family: Miridae Hahn, 1833

Genus: Grupocoris Douglas & Scott, 1868

Type species by monotypy: Grypocoris fieberi Douglas & Scott, 1868. (Ref.: Wagner, Loc. 1, 3♀♀ 2♂♂, 29.06.2017; Loc. 2, 1♀, 1966 (key).

Sub Genus: Turciocoris Wagner, 1966

Turciocoris Wagner, 1966:211 (as subgenus of Grypocoris).

Type species by original designation: Önder et al., 2006). Grupocoris Reuter, 1896. syriacus (Nomenclatural variant bv (1974): Grypocoris Turciocoris syriacus)

Grypocoris syriacus Reuter, 1896

Type locality: Akbez, (Hassa, Hatay)

Ent. Zeit. 15:256 (n.sp) Asia Minor, Syria 1896 Grypocoris syriacus Reuter, Hem. record (Aukema & Rieger, 1999). Gymn. Eur. 5:150 (descr., key)

1910 Grypocoris syriacus Oshanin, Verz. Pal. Hem.: 703 (cat) (Ref.:Carvalho, 1959) This species is an endemic species and

Habitat:

The individual specimens of this species was found in the scrub and meadow areas at an altitude of 1261-1344m.

Material examined: (Table 1)

30.06.2017.

Distribution in Turkey:

Adana, Osmaniye, Hatay, Kahramanmaraş (Reuter, 1896; Hoberlandt, 1956; Kezhner & Josifov, 1999; Aukema & Rieger, 1999;

With this study, this species was recorded Wagner for the first time from the north of Anatolia. This species is endemic to Anatolia

Distribution in Palaearctic

Turkey's Asian part (Anatolia), in the 1896 Grypocoris syriacus Reuter, Wien Palearctic catalog is in located Israel with a question mark (?) and it is a suspicious

Comments:

its type locality: Hatay (Hassa-Akbez).

According to Kerzner and Matocq (1994), Dursun, A. & Fent, M., 2017, Type Localities this species was first found in the Hatay (Hassa: Akbez) type locality by Reuter (1896) and later by Hoberlandt (1955) around Akbez (Adana, Osmaniye, Kahra- Hoberlandt, L., 1956, Results of the Zoological manmaras) in the Eastern Mediterranean region of Anatolia has mentioned.

Kerzner & Matocq (1994), referring to 2 specimens of the original Grypocoris syriacus Reuter, 1896 state that: possibly male and female, almost completely destroyed, only some legs and Kerzhner, I.M., 1997, Type specimens of some the apical part of hemelytra remaining.

Until this study, Grypocoris syriacus, which was endemic to the eastern and South-west Mediterranean region Turkey, appears to be limited to a small geographical area.

However, in the Palaeacrtic Catalougue, the distribution of this species in Anatolia, and in the as well as for Israel, is marked with a question mark and given as a suspicious record.

This study provides a better understanding of the distribution patterns of the species Reuter, O. M., 1896a, Dispositio generum in the Turkish Heteroptera fauna and the spread of G. syriacus to a new settlement area. Six rare examples G. syriacus Reuter, 1896 given in this study were obtained from Gerede (Bolu districts) in the Northern Anatolia Region of Turkey.

In order to understand the Turkish entomofauna, it helps to determine the entomgeographic features that affect the distribution of the fauna.

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A Preliminary List on the Host Plants of Lygaeoidea Species (Hemiptera: Heteroptera) in Turkey- II

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ABSTRACT: This study constitutes the second part of a preliminary list on the host plants of Lygaeoidea species in Turkey presented by Kıyak et al (2020).

In the first part of the study, Kıyak et al. (2020) presented and discussed the plant taxa belonging to 36 plant families and the related Lygaeoidea species list in alphabetical order.

In this study, 187 taxa belonging to 38 plant families, 67 at genera level and 120 at species level, were defined in relation to Lygaeoidea species. In addition, additional informations on the 10 plant families in the first study are presented.

KEYWORDS: Hemiptera, Lygaeoidea, host plant, relationship, Turkey.

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INTRODUCTION

The species belonging to the Lygaeoidea superfamily are phytophagous, except for the species of Geocoridae family, which are predators on various insect species. Information about the feeding habits of these species is given in the first part of the study (Kıyak et al., 2020).

literature data of previous years and studies conducted in Turkey in recent years.

genera level and 145 at the species level, were identified. belonging to 36 plant families were given between 1922 and 2020 (Kıyak et al., 2020).

In the last studies, it is seen that more first part of the study. animal and insect species have started to be seen outside their natural ranges due to many reasons such as increased human activation and global warming.

Some of these alien species increase excessively and reach a level that causes significant damage.

Heteroptera and even Lygaeoidea species.

Lygaeoidea species in Turkey, the both and the numbers of Lygaeoidea species studies aim i) to provide data about the related to them, and in Table 3 and food preferences of Lygaeoidea species Figure 2, plant families and plant taxon and the plant species and families to numbers belonging to these families are which they are more prone ii) to assist given. new studies on this group.

MATERIAL AND METHODS

The previous and recent studies on parentheses. Lygaeoidea species in Turkev were reviewed, and host plant lists were **RESULTS** created from these studies and listed according to plant families.

Fahringer Seidenstücker (1958,1963, Tuatay et al. (1972); Aysev (1974), Lodos added to the under of the table. et al. (1978, 1999), Çakır (1988), Özbek &

Alaoğlu (1988), Çakır & Önder (1990), Kıyak (1990, 1993, 2019), Çağlar (1992) Stichel (1957-1962) Özsaraç & Kıyak (2001), Özsaraç et al. (2001); Beyaz & Tezcan (2002), Atlihan et al. (2003), Gençer et al. (2004), Öncül Abacıgil et al. (2010), Tezcan et al. (2010), Matocq et al. (2014), Yıldırım & Eroğlu (2015), Dirik & Kıvan (2016), Kıyak & Özdamar (2017), In the first part of the study and in the Arslangundoğdu et al. (2018), Yücel & current study, a list of plants related to Kıvan (2018), Yazıcı (2019), Yazıcı & Sert-Lvgaeoidea species was created on the kava (2020), Cerci & Tezcan (2020), Özgen et al. (2020).

As a result of the literature review, plant taxa associated with Lygaeoidea species In the first part of the study, a total of in Turkey were listed according to 222 taxa, 77 of which were defined at the families, and a total of 74 plant families

> The first 36 of these plant families in alphabetical order and their related Lygaeoidea species were presented in the

> The remaining 38 families and their related taxa and related Lygaeoidea species are presented in Table 1 in alphabetical order in the present part of the study.

In addition. the missing records belonging to the plant families in the first These alien and invasive species include part of the study are added to the bottom of Table 1.

By making a list of plants related to In Table 2 and Figure 1 plant families

Plant species are given with their valid names, and the synonymous names in the old literature are also indicated in

In this part of the study, 187 plant taxa belonging to 38 plant families and related The literatures used in this study are; Lygaeoidea species are listed alphabetically (1922), Hoberlandt (1956), in Table 1. In addition, the additional 1965), records that were overlooked in Part I are

Table 1.The plant relationships of Lygaeoidea species in Turkey.

Lamiaceae		
Lamiaceae	Horvathiolus guttatus (Rambur, 1839)	Kıyak, 1993
	Metopoplax origani (Kolenati, 1845)	Lodos et al., 1999
Ajuga sp.	Pionosomus varius (Wolff, 1804)	Stichel, 1957
Ballota sp.	Beosus quadripunctatus (Müller, 1766) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Plinthisus longicollis Fieber, 1861	Lodos et al.,1999
Calamintha sp.	Ortholomus punctipennis (Herrich-Schaeffer, 1838)	Stichel,1957
Lavandula multifida L.	Heterogaster artemisiae Schilling, 1829	Stichel, 1957
Lycopus europaeus L.	Heterogaster cathariae (Geoffroy, 1785)	Stichel,1957
Marrubium parviflorum Fish & Mey.	Melanocoryphus albomaculatus (Goeze, 1778)	Kıyak 1990
Melissa officinalis L.	Heterogaster cathariae (Geoffroy, 1785)	Stichel,1957
Mentha sp.	Geocoris arenarius (Jakovlev, 1867) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Heterogaster artemisiae Schilling, 1829 Microplax interrupta (Fieber, 1837) Nysius cymoides (Spinola, 1837) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Scolopostethus affinis (Schilling, 1829)	Stichel,1957 Çakır,1988 Lodos et al., 1978, 1999
Mentha arvensis L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır,1988
Mentha piperita L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır,1988
Mentha rotundifolia (L.) Huds.	Oxycarenus pallens (Herrich-Schaeffer, 1850)	Aysev, 1974
Micromeria varia Benth.	Heterogaster artemisiae Schilling, 1829	Stichel, 1957
Nepeta sp.	Scolopostethus affinis (Schilling, 1829)	Stichel, 1957
Nepeta troodi Holmboe	Heterogaster cathariae (Geoffroy, 1785)	Stichel, 1957
Origanum sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Nysius cymoides (Spinola, 1837) Nysius graminicola graminicola (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius thymi thymi (Wolff, 1804)	Beyaz & Tezcan, 2002
Origanum syriacum L.	Geocoris luridus luridus (Fieber, 1844)	Çakır,1988
Origanum vulgare L.	Metopoplax origani (Kolenati, 1845)	Stichel 1957; Aysev, 1974
<i>Phlomi</i> s sp.	Artheneis foveolata Spinola, 1837 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius thymi thymi (Wolff, 1804) Oxycarenus pallens (Herrich-Schaeffer, 1850)	Çakır,1988; Lodos et al.,1999
Rosmarinus officinalis L.	Tropidothorax leucopterus (Goeze, 1778)	Stichel, 1957; Aysev, 1974
Salvia sp.	Emblethis ciliatus Horváth, 1875 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Graptopeltus lynceus (Fabricius, 1775) Heterogaster artemisiae Schilling, 1829 Microplax interrupta (Fieber, 1837) Nysius cymoides (Spinola, 1837) Nysius ericae ericae (Schilling, 1829) Nysius thymi thymi (Wolff, 1804) Platyplax salviae (Schilling, 1829)	Stichel, 1957; Ça- kır,1988; Lodos et al.,1999
Salvia glutinosa L.	Spilostethus saxatilis (Scopoli, 1763)	Stichel, 1957; Aysev,1974
Salvia pratensis L.	Platyplax salviae (Schilling, 1829)	Stichel, 1957
Salvia verticillata L.	Emblethis verbasci (Fabricius, 1803) Platyplax salviae (Schilling, 1829)	Stichel, 1957
Sideritis sp.	Geocoris ater (Fabricius, 1787)	Çakır,1988
Sideritis hyssopifolia L.	Cymus claviculus (Fallén, 1807) Ischnopeza hirticornis (Herrich-Schaeffer, 1850) Platyplax inermis (Rambur, 1839)	Lodos et al., 1978

	To it at the second of the 1760	Ö + 1 0001
Stachys cretica L.	Spilostethus saxatilis (Scopoli, 1763)	Özsaraç et al., 2001
Teucrium sp.	Emblethis verbasci (Fabricius, 1803) Geocoris erythrocephalus	Stichel, 1957 Özsaraç & Kıyak, 2001
Teucrium polium L.	<u> </u>	, , ,
Thymus sp.	Cymus claviculus (Fallén, 1807) Emblethis verbasci (Fabricius, 1803) Microplax interrupta (Fieber, 1837) Nysius graminicola graminicola (Kolenati, 1845) Ortholomus punctipennis (Herrich-Schaeffer, 1838) Pionosomus varius (Wolff, 1804)	Stichel, 1957; Aysev, 1974
Thymus serpyllum L.	Tropistethus holosericus (Scholtz, 1846)	Stichel, 1957
Thymus vulgaris L.	Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804)	Lodos et al., 1978, 1999
Lauraceae		
Laurus sp.	Heterogaster urticae (Fabricius, 1775) Metopoplax origani (Kolenati, 1845)	Lodos et al., 1999
Laurus nobilis L.	Spilostethus pandurus (Scopoli, 1763)	Lodos et al., 1999
Linaceae		
Linum sp.	Nysius ericae ericae (Schilling, 1829)	Stichel, 1957
Lythraceae		T
Lythrum portula (L.) D. A. Webb (Syn. Peplis portula L.)	Cymus claviculus (Fallén, 1807)	Stichel, 1957; Aysev, 1974
Punica sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır, 1988
Punica granatum L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844)	Çakır, 1988; Lodos et al., 1999
Malvaceae		
Abelmoschus esculentus (L.) Moench.	Oxycarenus hyalinipennis (A. Costa, 1843)	Lodos et al., 1999
(Syn. Hibiscus esculentus) Abutilon sp.	Oxycarenus hyalinipennis (A. Costa, 1843)	Stichel, 1957, Aysev,
noution sp.	Oxycurenus riguinipennus (ri. Costa, 1010)	1974
Althaea sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Oxycarenus hyalinipennis (A. Costa, 1843)	Aysev, 1974; Lodos et al., 1999
Althaea rosea Hohen. ex Boiss.	Oxycarenus hyalinipennis (A. Costa, 1843) Oxycarenus pallens (Herrich-Schaeffer, 1850) Metopoplax origani (Kolenati, 1845) Spilostethus pandurus (Scopoli, 1763)	Lodos et al., 1999
Gossypium sp.	Cymus melanocephalus Fieber, 1861 Geocoris arenarius (Jakovlev, 1867) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris lineola lineola (Rambur, 1839) Geocoris megacephalus (Rossi, 1790) Geocoris sp. Graptostetlzus servus (Fabricius, 1787) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus hyalinipennis (A. Costa, 1843)	Stichel, 1957; Tuatay et al., 1972; Aysev, 1974;Çakır, 1988; Çakır & Önder, 1990; Lodos et al.,1999; Yazıcı, 2019
Gossypium hirsutum L.	Leptodemus minutus Jakovlev 1876	Yazıcı & Sertkaya, 2020
Hibiscus sp.	Oxycarenus hyalinipennis (A. Costa, 1843)	Aysev, 1974
Malva sp.	Oxycarenus hyalinipennis (A. Costa, 1843)	Lodos et al., 1999
<i>Malva multiflora</i> (<u>Cav.</u>) Soldano, Banfi & Galasso	Oxycarenus hyalinipennis (A. Costa, 1843)	Stichel 1957; Aysev, 1974
(Syn. Lavatera cretica L.)		
Urena lobata L.	Graptostetlzus servus (Fabricius, 1787)	Stichel, 1957
Moraceae		
Morus sp.	Spilostethus pandurus (Scopoli, 1763)	Lodos et al.,1999
Morus alba L.	Megalonotus praetextatus (Herrich-Schaeffer, 1835) Spilostethus pandurus (Scopoli, 1763)	Kıyak, 1990; Lodos et al.,1999
Myrtaceae	1	I
Eucalyptus sp.	Horvathiolus superbus (Pollich, 1781)	Aysev, 1974
Nitrariaceae Peganum harmala L.	Emblethis ciliatus Horváth, 1875 Lygaeus equestris (Linnaeus, 1758) Tropidothorax leucopterus (Goeze, 1778)	Lodos et al., 1999

Oleaceae		
Fraxinus sp.	Caenocoris nerii (Germar, 1847) Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Aysev,1974; Çakır, 1988; Lodos et al.,1999
Fraxinus excelsior L.	Kleidocerys resedae resedae (Panzer, 1797) Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Aysev,1974; Çakır, 1988; Lodos et al., 1999
Jasminum fructicans L.	Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Oxycarenus pallens (Herrich-Schaeffer, 1850) Xanthochilus quadratus (Fabricius, 1798)	Çağlar, 1992
Ligustrum sp.	Geocoris luridus luridus (Fieber, 1844)	Çakır, 1988; Lodos et al.,1999
Olea europaea L.	Aphanus rolandri (Linnaeus, 1758) Arlheneis foveolata Spinola, 1837 Caenocoris nerii (Germar, 1847) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris lineola (Rambur, 1839) Geocoris megacephalus (Rossi, 1790) Graptostethus servus (Fabricius, 1787) Heterogaster urticae (Fabricius, 1775) Horvathiolus superbus (Pollich, 1781) Kleidocerys ericae (Horváth, 1908) Lamprodema maura (Fabricius, 1803) Lygaeus creticus (Lucas, 1854) Lygaeus equestris (Linnaeus, 1758) Macropternella inermis (Fieber, 1851) Metopoplax origani (Kolenati, 1845) Microplax albofasciata (A. Costa, 1847) Microplax limbata Fieber, 1864 Nysius cymoides (Spinola, 1837) Oxycarenus pallens (Herrich-Schaeffer, 1850) Peritrechus meridionalis Puton, 1877 Plinthisus longicollis Fieber, 1861 Proderus belloveyei Puton, 1874 Raglius alboacuminatus (Goeze, 1778) Remaudiereana annulipes (Bärensprung, 1859) Scolopostethus pictus (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763) Taphropeltus nervosus (Fieber, 1861) Xanthochilus quadratus (Fabricius, 1798)	Lodos et al.,1978, 1999; Çakır, 1988; Öncül Aba- cıgil et al., 2010
Onagraceae		
Epilobium hirsutum L.	Geocoris megacephalus (Rossi, 1790)	Stichel,1957; Aysev, 1974
Papaveraceae		
Papaveraceae	Brachyplax tenuis (Mulsant & Rey, 1852)	Matocq et al., 2014
Eschscholzia sp.	Spilostethus pandurus (Scopoli, 1763)	Lodos et al.,1999
Fumaria asapela Boiss.	Spilostethus pandurus (Scopoli, 1763)	Kıyak,1990
Fumaria judaica Boiss	Splilostethus pandurus (Scopoli, 1763)	Özsaraç & Kıyak, 2001
Papaver sp.	Brachyplax tenuis (Mulsant & Rey, 1852)	Seidenstücker, 1958
Pedaliaceae		
Sesamum indicum L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Nysius graminicola graminicola (Kolenati, 1845) Nysius senecionis senecionis (Schilling, 1829)	Lodos et al., 1978, 1999; Çakır,1988
Pinaceae		
Abies sp.	Spilostethus pandurus (Scopoli, 1763)	Lodos et al., 1999
Abies alba Mill.	Kleidocerys resedae resedae (Panzer, 1797)	Aysev, 1974
Cedrus sp.	Caenocoris nerii (Germar, 1847) Gastrodes grossipes (De Geer,1773) Geocoris ater (Fabricius, 1787) Geocoris luridus luridus (Fieber, 1844) Heterogaster artemisiae Schilling, 1829 Lasiocoris crassicornis (Lucas, 1849) Lygaeosoma sardeum erythropterum (Puton, 1876) Microplax limbata Fieber, 1864 Orsillodes longirostris Puton, 1884	Çakır, 1988; Lodos Önder et al 1999; Çerçi & Tez- can, 2020
Pinus sp.	Emblethis ciliatus Horváth, 1875 Gastrodes grossipes (De Geer,1773) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Graptopeltus lynceus (Fabricius, 1775) Heterogaster affinis Herrich-Schaeffer, 1835 Ischnopeza hirticornis (Herrich-Schaeffer, 1850)	Stichel, 1957; Çakır, 1988; Lodos et al., 1978, 1999

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Spilostethus saxatilis (Scopoli, 1763)	Pinus nigra J.F. Arnold	Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Megalonotus longipilis (Puton, 1884) Metopoplax fuscinervis Stål, 1872 Metopoplax origani (Kolenati, 1845) Nysius graminicola graminicola (Kolenati, 1845) Orsillus depressus (Mulsant & Rey, 1852) Orsillus reyi Puton, 1871 Plinthisus longicollis Fieber, 1861 Rhyparochromus vulgaris (Schilling, 1829) Scolopostethus affinis (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763) Spilostethus saxatilis (Scopoli, 1763) Graptopeltus validus (Horváth, 1875) Heterogaster affinis Herrich-Schaeffer, 1835 Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Ortholomus punctipennis (Herrich-Schaeffer, 1838) Oxycarenus pallens (Herrich-Schaeffer, 1850)	
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Pinus sylvestris L. Eremocoris plebejus (Fallen, 1807) Stichel, 1957; Yıldırım & Eroğlu, 2015	Pinus strobus L.	Eremocoris plebejus (Fallén, 1807)	Stichel, 1957
Plantaginaceae Antirhinum majus L. Nysius ericae ericae (Schilling, 1829) Misopates orontium (L.), Raf. (Syn. Antirhinum orontium) Plantago coronopus L. Henestaris laticeps laticeps (Curtis, 1836) Henestaris halophilus (Burmeister, 1835) Plantago indica L. Henestaris halophilus (Burmeister, 1835) Veronica sp. Beosus quadripunctatus (Müller, 1766) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Platanaceae Platanus sp. Geocoris luridus luridus (Fieber, 1844) Lygaeus equestris (Linnaeus, 1758) Megalonotus sabulicola (Thomson, 1870) Onder et al., 1999 Platanus orientalis L. Arocatus longiceps Stâl, 1872 Emblethis ciliatus Horváth, 1875 Geocoris luridus quadratus (Fieber, 1844) Spilostethus pandurus (Scopoli, 1763) Plumbaginaceae Acantholimon sp. Xanthochilus quadratus (Fabricius, 1798) Kyak, 1990 Poaceae Poaceae Cymophyes ochroleuca Fieber, 1870 Emblethis ciliatus Horváth, 1875 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Motopoplaya fasciata fasciata (Hersich-Schaeffer, 1849) Spilostethus pandurus (Scopoli, 1763) Plumbaginaceae Cymophyes ochroleuca Fieber, 1870 Emblethis ciliatus Horváth, 1875 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Motopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus pini (Unnaeus, 1758)	Pinus sylvestris L.	Eremocoris plebejus (Fallén, 1807)	
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Poaceae Poaceae Cymophyes ochroleuca Fieber, 1870 Emblethis angustus Montandon, 1890 Emblethis ciliatus Horváth, 1875 Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus hyalinipennis (A. Costa, 1843) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus pini (Unnaeus, 1758) Kiyak, 1990 Hoberlandt, 1956; Stichel, 1957; Ay- sev, 1974; Lodos Önder et al., 1978, 1999; Çakır, 1988 Cyakır, 1988		Emblethis ciliatus Horvath, 1875 Geocoris luridus luridus (Fieber, 1844)	
Poaceae Cymophyes ochroleuca Fieber, 1870 Hoberlandt, 1956; Stichel, 1957; Ay-Emblethis ciliatus Horváth, 1875 Stichel, 1957; Ay-Sev, 1974; Lodos Önder et al., 1978, 1999; Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus hyalinipennis (A. Costa, 1843) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus pini (Unnaeus, 1758)	Plumbaginaceae		
Poaceae Cymophyes ochroleuca Fieber, 1870 Emblethis angustus Montandon, 1890 Emblethis ciliatus Horváth, 1875 Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus hyalinipennis (A. Costa, 1843) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus pini (Unnaeus, 1758)	<u>-</u>	Xanthochilus quadratus (Fabricius, 1798)	Kıyak, 1990
Emblethis angustus Montandon, 1890 Emblethis ciliatus Horváth, 1875 Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus hyalinipennis (A. Costa, 1843) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus pini (Unnaeus, 1758)	Poaceae		
Spilostethus pandurus (Scopoli, 1763) Spilostethus saxatilis (Scopoli, 1763)	Poaceae	Emblethis angustus Montandon, 1890 Emblethis ciliatus Horváth, 1875 Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Henestaris halophilus (Burmeister, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus hyalinipennis (A. Costa, 1843) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus pini (Unnaeus, 1758)	Stichel, 1957; Aysev,1974; Lodos Önder et al., 1978, 1999;
Applions caudata I. I Bensus maritimus (Scopoli 1763) Ozsarac et al. 2001		Spilostethus saxatilis (Scopoli, 1763)	B
Agropuron sp. Nysius cymoides (Spinola, 1837) Lodos et al., 1999	Aegilops caudata L.	Spilostethus saxatilis (Scopoli, 1763) Beosus maritimus (Scopoli, 1763)	Özsaraç et al., 2001
Applions caugata L. L. Beosus maritimus (Scopoli, 1763) L. Ozsarac et al., 2001	A '1 1 4 7	Spilostethus saxatilis (Scopoli, 1763)	Ö 1 . 2001

Andropogon dystachius L.	Paromius gracilis (Rambur, 1839)	Stichel, 1957
Avena barbata Pott ex Link	Oxycarenus pallens (Herrich-Schaeffer, 1850)	Aysev, 1974
Bromus arvensis L.	Beosus maritimus (Scopoli, 1763)	Özsaraç et al., 2001
Bromus madritensis L.	Metopoplax fuscinervis Stål, 1872	Özsaraç et al., 2001
Elytrigia repens (L.) Newski (Syn. Agropyron repens)	Paromius gracilis (Rambur, 1839)	Lodos et al., 1999
Glyceria sp.	lschnodemus suturalis Horváth, 1883	Seidenstücker, 1958
Imperata arundinacea Cirillio	Paromius gracilis (Rambur, 1839)	Stichel, 1957
Hordeum sp.	Oxycarenus pallens (Herrich-Schaeffer, 1850) Spilostethus saxatilis (Scopoli, 1763)	Lodos et al., 1999
Hordeum vulgare L.	Spilostethus pandurus (Scopoli, 1763)	Lodos et al.,1999
Lolium sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır, 1988
Oryza sativa L.	Geocoris megacephalus (Rossi, 1790)	Çakır, 1988
Panicum turgidum Forrsk.	Cymophyes ochroleuca Fieber, 1870	Stichel, 1957
Phragmites sp.	Cymus melanocephalus Fieber, 1861 Dimorphopterus blissoides (Baerensprung, 1859)	Stichel, 1957; Lodos et al., 1999
Poa bulbosa var. vivipara Koeler	Spilostethus pandurus (Scopoli, 1763)	Kıyak, 1990
Saccharum revanne L. (Syn. Erianthus ravennae)	Paromius gracilis (Rambur, 1839)	Stichel, 1957
Secale cereale L.	Aphanus rolandri (Linnaeus, 1758)	Lodos et al., 1999
Sorghum halepense (L.) Pers.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır, 1988
Triticum sp. Triticum aestivum L.	Beosus maritimus (Scopoli, 1763) Cymus claviculus (Fallén, 1807) Emblethis denticollis Horváth, 1878 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Heterogaster urticae (Fabricius, 1775) Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Megalonotus praetextatus (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus modestus (Fallén, 1829) Oxycarenus pallens (Herrich-Schaeffer, 1850) Rhyparochromus phoeniceus (Rossi, 1794) Rhyparochromus vulgaris (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778) Xanthochilus satumius (Rossi, 1790) Spilostethus pandurus (Scopoli, 1763)	Lodos et al., 1978,1999; Çakır, 1988; Dirik & Kıvan, 2016 Alkan, 1948; Lodos et
(Syn. Triticum vulgare, Triticum sativum)	Spilostethus saxatilis (Scopoli, 1763) Metopoplax fuscinervis Stål, 1872	al., 1999; Özsaraç et al., 2001
Zea mays L.	Nysius cymoides (Spinola, 1837) Scolopostethus pictus (Schilling, 1829)	Lodos et al., 1978; Özgen et al., 2020
Polygonaceae		,
Atraphaxis sp.	Emblethis setifer Seidenstücker, 1966	Seidenstücker, 1967
Polygonum sp.	Metopoplax origani (Kolenati, 1845) Oxycarenus pallens (Herrich-Schaeffer, 1850)	Aysev, 1974
Rumex sp.	Geocoris megacephalus (Rossi, 1790) Heterogaster artemisiae Schilling, 1829 Lygaeus equestris (Linnaeus, 1758) Xanthochilus minusculus (Reuter, 1885)	Kıyak, 1993; Çakır, 1988; Lodos et al., 1999
Rumex acetosella L.	Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845)	Lodos et al., 1978
Rumex scutatus L.	Horvathiolus superbus (Pollich, 1781)	Stichel, 1957
Portulacaceae		
Portulaca sp.	Geocoris megacephalus (Rossi, 1790) Geocoris lineola lineola (Rambur, 1839)	Aysev, 1974
Portulaca oleracea L.	Nysius senecionis senecionis (Schilling, 1829)	Fahringer, 1922; Hoberlandt, 1956; Stichel, 1957

Ranunculaceae		
Adonis sp.	Lygaeus equestris (Linnaeus, 1758)	Stichel, 1957; Aysev, 1974
Nigella arvensis L.	Apterola lowni (Saunders, 1876)	Kıyak & Özdamar, 2017
Nigella sativa L.	Lygaeus equestris (Linnaeus, 1758)	Stichel, 1957; Aysev, 1974
Rhamnaceae		
Paliurus sp.	Orsillus depressus (Mulsant & Rey, 1852)	Lodos et al.,1999
Paliurus spina-christi Mill.	Caenocoris nerii (Germar, 1847) Lygaeus creticus Lucas, 1854	Lodos et al., 1978
	Lygaeus equestris (Linnaeus, 1758)	
Rosaceae		
Amygdalus sp.	Brachyplax tenuis (Mulsant & Rey, 1852) Oxycarenus pallens (Herrich-Schaeffer, 1850)	Lodos et al., 1999
Amygdalus communis L. (Syn. Prunus amygdalus)	Emblethis denticollis Horváth, 1878 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Kleidocerys ericae (Horváth, 1908) Lygaeus creticus Lucas, 1854 Lygaeus equestris (Linnaeus, 1758) Megalonotus sabulicola (Thomson, 1870) Metopoplax origani (Kolenati, 1845) Spilostethus pandurus (Scopoli, 1763)	Lodos et al., 1978, 1999; Çakır, 1988
Aruncus sylvester Kostel ex Mexim	Spilostethus saxatilis (Scopoli, 1763)	Stichel, 1957
Cotoneaster integerrimus Medik.	Horvathiolus superbus (Pollich, 1781)	Stichel, 1957; Aysev, 1974
Crataegus sp.	Arocatus melanocephalus (Fabricius, 1798) Artheneis alutacea Fieber, 1861 Emblethis denticollis Horváth, 1878 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Heterogaster urticae (Fabricius, 1775) Metopoplax origani (Kolenati, 1845) Nysius graminicola graminicola (Kolenati, 1845) Spilostethus pandurus (Scopoli, 1763)	Stichel, 1957; Çakır, 1988; Lodos et al.,1999
Cydonia oblanga Mill. (Syn. Cydonia vulgaris)	Beosus maritimus (Scopoli, 1763) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Lygaeus equestris (Linnaeus,1758) Metopoplax origani (Kolenati, 1845) Raglius zarudnyi (Jakovlev, 1905) Rhyparochromus vulgaris (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763)	Lodos et al.,1978, 1999; Gençer et al., 2004
Fragaria sp.	Nysius ericae ericae (Schilling, 1829)	Stichel, 1957
Fragaria vesca L.	Emblethis angustus Montandon, 1890 Geocoris megacephalus (Rossi, 1790)	Lodos et al., 1978; Yazıcı, 2019
Malus angustifolia (Aiton) Michx. (Syn. Pyrus angustifolia)	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Lodos et al., 1999
Malus domestica Borkh. (Syn. Malus communis, Pyrus malus)	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Kleidocerys resedae resedae (Panzer, 1797) Lygaeus creticus Lucas, 1854 Lygaeus equestris (Linnaeus, 1758) Metopoplax fuscinervis Stål, 1872 Oxycarenus pallens (Herrich-Schaeffer, 1850) Peritrechus meridionalis Puton, 1877 Scolopostethus affinis (Schilling, 1829) Spilostethus saxatilis (Scopoli, 1763)	Aysev, 1978; Lodos et al., 1978, 1999
Malus sylvestris (L.) Mill.	Emblethis griseus (Wolff, 1802) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Lygaeus creticus Lucas, 1854 Lygaeus equestris (Linnaeus, 1758) Nysius graminicola graminicola (Kolenati, 1845) Spilostethus pandurus (Scopoli, 1763)	Yiğit & Uygun, 1982
Mespilus germanica L.	Heterogaster urticae (Fabricius, 1775) Lygaeus creticus Lucas, 1854	Lodos et al., 1978
Pirus elaeagrifolia Pall.	Geocoris ater (Fabricius, 1787) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Taphropeltus nervosus (Fieber, 1861) Xanthochilus minusculus (Reuter, 1885)	Lodos et al., 1978

Potentilla sp.	Tropistethus holosericus (Scholtz, 1846)	Stichel, 1957
Potentilla neumanniana Rchb. (Syn. Potentilla verna)	Ortholomus punctipennis (Herrich-Schaeffer, 1838)	Stichel, 1957; Aysev, 1974
Prunus sp.	Leptodemus minutus (Jakovlev, 1874) Stygnocoris rusticus (Fallén, 1807)	Lodos et al., 1978, 1999
Prunus armeniaca L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster artemisiae Schilling, 1829 Heterogaster urticae (Fabricius, 1775) Lygaeus equestris (Linnaeus, 1758) Peritrechus meridionalis Puton, 1877 Spilostethus saxatilis (Scopoli, 1763)	Çakır, 1988; Lodos et al., 1999
Prunus avium L.	Aphanus rolandri (Linnaeus, 1758) Taphropeltus nervosus (Fieber, 1861) Beosus quadripunctatus (Müller 1766) Eremocoris fenestratus (Herrich & Schaeffer 1839) Graptostethus servus (Fabricius 1787) Icus angularis Fieber 1861 Lamprodema maura (Fabricius 1803) Nysius graminicola graminicola (Kolenatı 1845) Nysius helveticus (Herrich & Schaeffer 1850) Paromius gracilis (Rambur 1839) Remaudiereana annulipes (Barensprung 1859) Scolopostethus pictus (Schilling 1829)	Lodos et al., 1978, 1999; Tezcan et al., 2010
Prunus cerasus L.	Metopoplax origani (Kolenati, 1845)	Lodos et al.,1978
Prunus domestica L.	Beosus maritimus (Scopoli, 1763) Geocoris luridus luridus (Fieber, 1844) Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Lygaeus equestris (Linnaeus, 1758) Megalonotus sabulicola (Thomson, 1870) Metopoplax origani (Kolenati, 1845)	Lodos et al., 1978, 1999; Çakır, 1988
Prunus mahaleb L.	Lygaeus creticus Lucas, 1854	Lodos, Önder et al., 1978
Prunus persica (L.) Batsh.	Beosus maritimus (Scopoli, 1763) Lygaeosoma anatolicum Seidenstücker, 1960 Lygaeus creticus Lucas, 1854 Lygaeus equestris (Linnaeus, 1758)	Lodos et al., 1978, 1999
Pyrus communis L.	Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Nysius cymoides (Spinola, 1837) Peritrechus meridionalis Puton, 1877	Lodos et al., 1978, 1999
Pyrus elaeagrifolia Pall.	Geocoris ater (Fabricius, 1787) Heterogaster affinis Herrich-Schaeffer, 1835 Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Megalonotus praetextatus (Herrich-Schaeffer, 1835) Taphropeltus nervosus (Fieber, 1861) Xanthochilus minusculus (Reuter, 1885)	Lodos et al., 1978, 1999
Rosa sp.	Beosus maritimus (Scopoli, 1763) Engistus salinus (Jakovlev, 1874) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Lygaeus creticus Lucas, 1854 Lygaeus equestris (Linnaeus, 1758) Metopoplax origani (Kolenati, 1845) Oxycarenus hyalinipennis (A. Costa, 1843) Spilostethus pandurus (Scopoli, 1763) Stygnocoris rusticus (Fallén, 1807)	Lodos et al.,1978, 1999; Çakır, 1988; Yücel & Kıvan, 2018
Rosa canina L.	Eremocoris praenotatus Seidenstücker, 1965	Seidenstücker, 1965
Rubus sp.	Arocatus longiceps Stål, 1872 Beosus maritimus (Scopoli, 1763) Cymus melanocephalus Fieber, 1861 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Kleidocerys ericae (Horváth, 1908) Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Oxycarenus pallens (Herrich-Schaeffer, 1850) Scolopostethus pictus (Schilling, 1829)	Lodos, Önder et al., 1978, 1999; Çakır, 1988
Rubus ideaus L.	Spilostethus pandurus (Scopoli, 1763)	Kıyak, 1990

Sanguisorba minor Scop.	Emblethis dilaticollis (Jakovlev, 1874) Ortholomus jordani Hoberlandt, 1953 Raglius confusus (Reuter, 1886)	Kıyak, 1990; Lodos et al., 1999
Sanguisorba officinalis L.	Spilostethus saxatilis (Scopoli, 1763)	Stichel, 1957; Aysev, 1974
Sarcopterium spinosum (L.) Spach.	Macroplax fasciata fasciata (Herrich-Schaeffer, 1835)	Özsaraç et al., 2001
Sorbus aria (L.) Crantz.	Lygaeus creticus Lucas, 1854	Stichel, 1957; Aysev, 1974
Rubiaceae		
Galium sp.	Cymus claviculus (Fallén, 1807)	Stichel, 1957; Aysev, 1974
Galium mollugo L.	Spilostethus saxatilis (Scopoli, 1763)	Stichel, 1957; Aysev, 1974
Rutaceae		
Citrus sp.	Metopoplax fuscinervis Stål, 1872 Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Heterogaster urticae (Fabricius, 1775)	Lodos et al., 1978,1999
Citrus unshui Marc.	Lygaeus creticus Lucas, 1854	Lodos et al., 1978
Salicaceae		
Populus sp.	Arlheneis foveolata Spinola, 1837 Geocoris erythrocephalus (Lepeletier & Serville, 1825) Gonianotus galactodennus Fieber, 1861 Graptopeltus validus (Horváth, 1875) Homoscelis ruficollis Horváth, 1884 Leptodemus minutus (Jakovlev, 1874) Lygaeus equestris (Linnaeus, 1758) Ortholomus carinatus (Lindberg, 1932) Spilostethus pandurus (Scopoli, 1763)	Stichel, 1957; Seidens- tücker, 1960; Lodos et al., 1978, 1999; Çakır, 1988
Populus alba L.	Leptodemus minutus (Jakovlev, 1874)	Lodos et al., 1978
Populus nigra L.	Kleidocerys resedae resedae (Panzer, 1797)	Aysev, 1974; Yıldırım & Eroğlu, 2015
Populus tremula L.	Artheneis alutacea Fieber, 1861	Lodos et al., 1978
Salix sp.	Artheneis alutacea Fieber, 1861 Arlheneis foveolata Spinola, 1837 Arlheneis hyrcanica (Kolenati, 1845) Artheneis balcanica (Kormilev, 1938) Beosus maritimus (Scopoli, 1763) Caenocoris nerii (Germar, 1847) Camptocera glaberrima (Walker, 1872) Cymus glandicolor Hahn, 1832 Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Horvathiolus superbus (Pollich, 1781) Leptodemus minutus (Jakovlev, 1874) Lygaeus equestris (Linnaeus, 1758) Metopoplax origani (Kolenati, 1845) Microplax limbata Fieber, 1864 Oxycarenus pallens (Herrich-Schaeffer, 1850) Peritrechus meridionalis Puton, 1877 Scolopostethus decoratus (Hahn, 1833) Spilostethus saxatilis (Scopoli, 1763)	Stichel, 1957; Aysev, 1974; Lodos et al., 1978, 1999; Çakır, 1988; Matocq et al., 2014; Kıyak, 2019
Salix aurita L.	Kleidocerys resedae resedae (Panzer, 1797) Kleidocerys resedae resedae (Panzer, 1797)	Aysev, 1974
Salix babylonica L. Santalaceae	memocerys reseaue reseaue (ranzer, 1797)	Yıldırım & Eroğlu, 2015
Viscum album L.	Spilostethus saxatilis (Scopoli, 1763)	Lodos et al., 1999
Scrophulariaceae	Spassenus saxams (Scopon, 1703)	2000s ct al., 1399
•	Hotorogastar artemisiae Schilling 1900	Aveev 1074: Kwelz
Scrophularia sp.	Heterogaster artemisiae Schilling, 1829 Nysius graminicola graminicola (Kolenati, 1845) Oxycarenus pallens (Herrich-Schaeffer, 1850) Spilostethus pandurus (Scopoli, 1763)	Aysev, 1974; Kıyak, 1990; Lodos et al., 1999
Verbascum sp.	Aphanus rolandri (Linnaeus, 1758) Arlheneis foveolata Spinola, 1837 Cymus melanocephalus Fieber, 1861 Emblethis angustus Montandon, 1890 Emblethis ciliatus Horváth, 1875 Emblethis verbasci (Fabricius, 1803) Eremocoris fenestratus (Herrich-Schaeffer, 1839)	Stichel, 1957; Lodos et al., 1978, 1999; Aysev, 1974; Kıyak, 1993; Ça- kır, 1988; Çakır, & Ön- der, 1990;

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Gastrodes grossipes (De Geer, 1773) Geocoris arenarius (Jakovlev, 1867) Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Geocoris megacephalus (Rossi, 1790) Heterogaster urticae (Fabricius, 1775) Horvathiolus superbus (Pollich, 1781) Lamprodema maura (Fabricius, 1803) Lygaeosoma anatolicum Seidenstücker, 1960 Lygaeosoma sardeum sardeum Spinola, 1837 Lygaeus equestris (Linnaeus, 1758)	Stichel, 1957; Lodos et al., 1978, 1999; Aysev, 1974; Kıyak, 1993; Çakır, 1988; Çakır, & Önder, 1990;
Macroplas Jasciata Jasciata (Herrich-Schaeffer, 1835) Megalonotus emarginatus (Rey, 1888) Megalonotus sabulicola (Thomson, 1870) Metopoplax fuscinervis Stål, 1872 Metopoplax origani (Kolenati, 1845) Microplax interrupta (Fieber, 1837) Nvsius senecionis senecionis (Schilling, 1829) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus pallens (Herrich-Schaeffer, 1850) Plinthisus longicollis Fieber, 1861 Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus cognatus Fieber, 1861 Spilostethus pandurus (Scopoli, 1763)	
Spilostethus saxatilis (Scopoli, 1763)	
Xanthochilus minusculus (Reuter, 1885)	Öggggggggggggggggggggggggggggggggggggg
Еливетть angustus montandon, 1890	Özsaraç et al., 2001
Nucius aumaidas (Spinols, 1927)	Özgen et al., 2020
	Stichel, 1957; Kıyak,
Nysius cymoides (Spinola, 1837) Nysius ericae ericae (Schilling, 1829)	1990
Nysius cymoides (Spinola, 1837)	Özgen et al., 2020
Nysius cymoides (Spinola, 1837)	Özgen et al., 2020
Geocoris pallidipennis pallidipennis (A. Costa, 1843) Emblethis griseus (Wolff, 1802) Lygaeus equestris (Linnaeus, 1758) Nysius cymoides (Spinola, 1837) Nysius ericae ericae (Schilling, 1829) Oxucarenus pallens (Herrich-Schaeffer, 1850)	Stichel, 1957; Aysev, 1974; Özbek & Alaoğlu, 1988; Atlıhan et al., 2003; Özgen et al., 2020
Lygaeosoma sardeum sardeum Spinola, 1837	Lodos et al., 1999
•	
Arocatus melanocephalus (Fabricius, 1798) Caenocoris nerii (Germar, 1847) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Spilostethus pandurus (Scopoli, 1763)	Çakır, 1988; Lodos et al., 1999
Geocoris erythrocephalus (Lepeletier & Serville, 1825) Beosus maritimus (Scopoli, 1763)	Lodos et al., 1978
Artheneis alutacea Fieber, 1861	Lodos, Önder et al., 1999
Henestaris laticeps laticeps (Curtis, 1836)	Seidenstücker, 1958
Artheneis alutacea Fieber, 1861 Artheneis balcanica (Kormilev, 1938) Arlheneis foveolata Spinola, 1837 Arlheneis hyrcanica (Kolenati, 1845) Beosus quadripunctatus (Müller, 1766) Caenocoris nerii (Germar, 1847) Cymus melanocephalus Fieber, 1861 Emblethis brachynotus Horvath, 1897 Emblethis griseus (Wolff, 1802)	Hoberlandt, 1956; Stichel, 1957; Seidenstücker, 1958; Lodos et al., 1978, 1999; Çakır & Önder, 1990; Çakır,1988; Lodos et al., Matocq et al., 2014
	Geocoris arenarius (Jakovlev, 1867) Geocoris ater (Fabricius, 1787) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Geocoris megacephalus (Rossi, 1790) Heterogaster urticae (Fabricius, 1775) Horvathiolus superbus (Pollich, 1781) Lamprodema maura (Fabricius, 1803) Lygaeosoma anatolicum Seidenstücker, 1960 Lygaeosoma sardeum Seidenstücker, 1960 Lygaeosoma sardeum sardeum Spinola, 1837 Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Megalonotus emarginatus (Rey, 1888) Megalonotus sabulicola (Thomson, 1870) Metopoplax fuscineruis Stäl, 1872 Metopoplax fuscineruis Stäl, 1872 Metopoplax interrupta (Fieber, 1837) Nusius senecionis senecionis (Schilling, 1829) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus pallens (Herrich-Schaeffer, 1850) Plinthisus longicollis Fieber, 1861 Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus cognatus Fieber, 1861 Spilostethus pandurus (Scopoli, 1763) Spilostethus saxatilis (Scopoli, 1763) Spilostethus asynatilis (Scopoli, 1763) Spilostethus asynatilis (Scopoli, 1763) Xanthochilus minusculus (Reuter, 1885) Emblethis angustus Montandon, 1890 Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Nysius cymoides (Spinola, 1837) Ocycarenus pallens (Herrich-Schaeffer, 1850) Lygaeus equestris (Linnaeus, 1758) Nysius cymoides (Spinola, 1837) Nysius ericae ericae (Schilling, 1829) Oxycarenus pallens (Herrich-Schaeffer, 1850) Lygaeosoma sardeum sardeum Spinola, 1837 Arocatus melanocephalus (Fabricius, 1798) Caenocoris nerii (Germar, 1847) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Beosus maritimus (Scopoli, 1763) Lygaeus creticus Lucas, 1854 Artheneis alutacea Fieber, 1861 Henestaris laticeps laticeps (Curt

	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Holcocranum saturejae (Kolenati, 1845) Hyalochilus dolosus Horváth, 1897 Lygaeosoma sardeum erythropterum (Puton, 1876) Lygaeus equestris (Linnaeus, 1758) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Nysius ericae ericae (Schilling, 1829) Nysius graminicola graminicola (Kolenati, 1845) Nysius thymi thymi (Wolff, 1804) Oxycarenus pallens (Herrich-Schaeffer, 1850) Paromius gracilis (Rambur, 1839) Rhyparochromus vulgaris (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763)	
Thymelaeaceae		
Daphne gnidioides L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır, 1988
Tiliaceae		
Tilia sp.	Kleidocerys resedae resedae (Panzer, 1797) Arocatus melanocephalus (Fabricius, 1798)	Stichel, 1957; Aysev, 1974
Tilia tomentosa Moench	Oxycarenus lavaterae (Fabricius, 1787)	Arslangündoğdu et al., 2018
Typhaceae		L 1 1 1000
Typha sp.	lschnodemus caspius Jakovlev, 1871	Lodos et al.,1999
Ulmaceae		1
Ulmus minor Mill. (Syn. Ulmus campestris)	Caenocoris nerii (Germar, 1847)	Lodos et al.,1978
Ulmus sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Metopoplax origani (Kolenati, 1845) Oxycarenus pallens (Herrich-Schaeffer, 1850) Spilostethus saxatilis (Scopoli, 1763)	Çakır, 1988; Lodos et al.,1999
Ulmus glabra Huds.	Arocatus melanocephalus (Fabricius, 1798)	Stichel, 1957
Urticaceae		
Parietaria debilis G. Forst	Hyalochilus ovatulus (A. Costa, 1853)	Stichel, 1957
Parietaria vulgaris Hill	Hyalochilus dolosus Horváth, 1897	Stichel, 1957
Urtica en	Beosus maritimus (Scopoli, 1763) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stál, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus pictus (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778)	
Urtica sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stål, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus cognatus Fieber, 1861 Scolopostethus pictus (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778)	Hoberlandt, 1956; Stichel, 1957; Çakır, 1988; Lodos, Önder et al., 1999
Urtica dioeca L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stål, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus cognatus Fieber, 1861 Scolopostethus pictus (Schilling, 1829)	hel, 1957; Çakır, 1988; Lodos, Önder et al.,
	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stål, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus cognatus Fieber, 1861 Scolopostethus pictus (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778) Heterogaster urticae (Fabricius, 1775) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Microplax albofasciata (A. Costa, 1847) Nysius graminicola graminicola (Kolenati, 1845) Rhyparochromus vulgaris (Schilling, 1829)	hel, 1957; Çakır, 1988; Lodos, Önder et al., 1999 Stichel, 1957; Aysev,
Urtica dioeca L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stål, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus cognatus Fieber, 1861 Scolopostethus pictus (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778) Heterogaster urticae (Fabricius, 1775) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Microplax albofasciata (A. Costa, 1847) Nysius graminicola graminicola (Kolenati, 1845) Rhyparochromus vulgaris (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763)	hel, 1957; Çakır, 1988; Lodos, Önder et al., 1999 Stichel, 1957; Aysev, 1974; Lodos et al., 1978 Stichel, 1957; Aysev, 1974 Stichel, 1957; Aysev,
Urtica dioeca L. Urtica morifolia Poir.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stål, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus cognatus Fieber, 1861 Scolopostethus pictus (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778) Heterogaster urticae (Fabricius, 1775) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Microplax albofasciata (A. Costa, 1847) Nysius graminicola graminicola (Kolenati, 1845) Rhyparochromus vulgaris (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763) Heterogaster urticae (Fabricius, 1775)	hel, 1957; Çakır, 1988; Lodos, Önder et al., 1999 Stichel, 1957; Aysev, 1974; Lodos et al., 1978 Stichel, 1957; Aysev, 1974
Urtica dioeca L. Urtica morifolia Poir. Urtica urens L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Heterogaster affinis Herrich-Schaeffer, 1835 Heterogaster urticae (Fabricius, 1775) Kleidocerys ericae (Horváth, 1908) Metopoplax fuscinervis Stål, 1872 Orsillus depressus (Mulsant & Rey, 1852) Raglius pineti (Herrich-Schaeffer, 1835) Rhyparochromus phoeniceus (Rossi, 1794) Scolopostethus affinis (Schilling, 1829) Scolopostethus cognatus Fieber, 1861 Scolopostethus pictus (Schilling, 1829) Tropidothorax leucopterus (Goeze, 1778) Heterogaster urticae (Fabricius, 1775) Macroplax fasciata fasciata (Herrich-Schaeffer, 1835) Metopoplax origani (Kolenati, 1845) Microplax albofasciata (A. Costa, 1847) Nysius graminicola graminicola (Kolenati, 1845) Rhyparochromus vulgaris (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763) Heterogaster urticae (Fabricius, 1775)	hel, 1957; Çakır, 1988; Lodos, Önder et al., 1999 Stichel, 1957; Aysev, 1974; Lodos et al., 1978 Stichel, 1957; Aysev, 1974 Stichel, 1957; Aysev,

Vitex agnus-castus L	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Xanthochilus saturnius (Rossi, 1790)	Çakır, 1988, Lodos et al., 1999; Özsaraç et al., 2001	
Vitaceae			
Vitis vinifera L.	Heterogaster urticae (Fabricius, 1775) Lygaeus equestris (Linnaeus, 1758) Megalonotus sabulicola (Thomson, 1870) Metopoplax origani (Kolenati, 1845) Nysius cymoides (Spinola, 1837) Nvsius senecionis senecionis (Schilling, 1829) Nysius ericae ericae (Schilling, 1829) Platyplax salviae (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763)	Stichel, 1957; Lodos et al., 1978, 1999	
Additional Information for Part I (Kıyak et al, 2020).		
Apiaceae			
Apiaceae (Umbelliferae)	iaceae (Umbelliferae) Beosus maritimus (Scopoli, 1763) Geocoris erythrocephalus (Lepeletier & Serville, 1825) Geocoris luridus luridus (Fieber, 1844) Lygaeus equestris (Linnaeus, 1758) Nysius graminicola graminicola (Kolenati, 1845)		
Conium maculatum L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Yazıcı, 2019	
Apocynaceae			
Nerium oleander L.	Arocatus longiceps Stål, 1872 Caenocoris nerii (Germar, 1847) Lygaeus creticus Lucas, 1854 Lygaeus equestris (Linnaeus, 1758) Metopoplax origani (Kolenati, 1845) Scolopostethus pictus (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763) Tropidothorax leucopterus (Goeze, 1778)	Stichel, 1957; Aysev, 1974; Lodos et al., 1978, 1999	
Asparagaceae			
Asparagus sp.	Geocoris megacephalus (Rossi, 1790)	Aysev, 1974	
Asphodelaceae		•	
Asphodelus sp.	Beosus maritimus (Scopoli, 1763) Rhyparochromus vulgaris (Schilling, 1829) Spilostethus pandurus (Scopoli, 1763)	Lodos et al., 1999	
Asphodelus microcarpus Viv.	Macroplax fasciata fasciata (Herrich-Schaeffer, 1835)	Lodos et al, 1978	
Asteraceae			
Asteraceae (Compositae)	Plinthisus longicollis Fieber, 1861	Lodos et al., 1999	
Echinops sp.	Pionosomus persimilis Horváth, 1895	Hoberlandt, 1956	
Cannabaceae			
Cannabis sp.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Çakır, 1988; Lodos et al., 1999	
Cannabis sativa L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825) Kleidocerys ericae (Horváth, 1908) Spilostethus saxatilis (Scopoli, 1763)	Lodos et al., 1978, 1999	
Colchicaceae			
Colchicum autumnale L.	Spilostethus saxatilis (Scopoli, 1763)	Stichel, 1957; Aysev, 1974	
Cupressaceae			
Thuja sp.	uja sp. Orsillus maculatus (Fieber, 1861) Orsillus depressus (Mulsant & Rey, 1852)		
Platycladus orientalis L. (Syn. Thuja orientalis)	Orsillus depressus (Mulsant & Rey, 1852)	Stichel, 1957	
Ericaceae			
Rhododendron tomentosum Harma- ja (Syn. Ledum palustre)	Kleidocerys resedae resedae (Panzer, 1797)	Aysev, 1974	
Fabaceae			
Fabaceae (Leguminosae)	Cymus melanocephalus Fieber, 1861 Oxycarenus pallens (Herrich-Schaeffer, 1850) Rhyparochromus phoeniceus (Rossi, 1794) Stygnocoris sabulosus (Schilling, 1829)	Lodos et al., 1999	
Acacia sp. Geocoris luridus (Fieber, 1844) Nysius cymoides (Spinola, 1837) Plinthisus mehadiensis Horvath, 1881		Lodos et al.,1999; Çakır & Önder, 1990	
Medicago sativa L.	Geocoris erythrocephalus (Lepeletier & Serville, 1825)	Yazıcı, 2019	

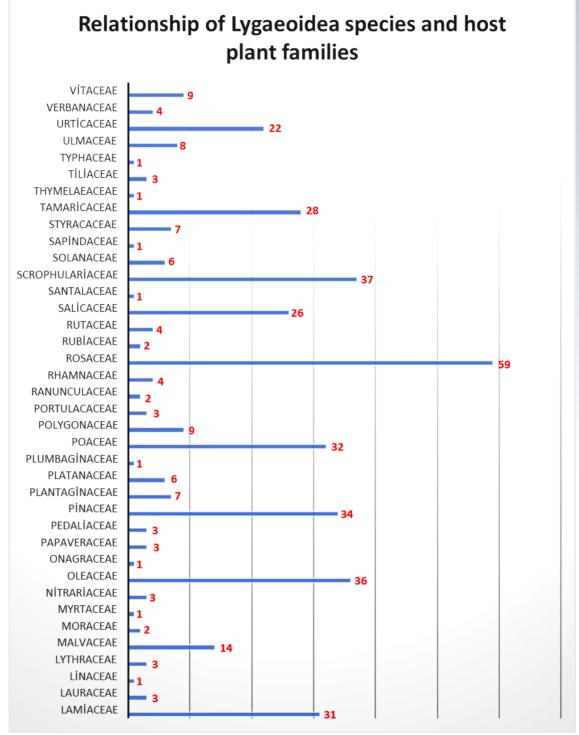


Figure 1. Distribution of Lygaeoidea species numbers according to families of host plants in Turkey.

Table 2. The number of Lygaeoidea species on hostplants family in Turkey

Host plant Fa- mily	Number of Lygaeoidea species on the host	Host plant Family	Number of Lygaeoidea species on the host	
Lamiaceae	31	Ranunculaceae	2	
Lauraceae	3	Rhamnaceae	4	
Linaceae	1	Rosaceae	59	
Lythraceae	3	Rubiaceae	2	
Malvaceae	14	Rutaceae	4	
Moraceae	2	Salicaceae	26	
Myrtaceae	1	Santalaceae	1	
Nitrariaceae	3	Scrophulariaceae	37	
Oleaceae	36	Solanaceae	6	
Onagraceae	1	Sapindaceae	1	
Papaveraceae	3	Styracaceae	7	
Pedaliaceae	3	Tamaricaceae	28	
Pinaceae	34	Thymelaeaceae	1	
Plantaginaceae	7	Tiliaceae	3	
Platanaceae	6	Typhaceae	1	
Plumbaginaceae	1	Ulmaceae	8	
Poaceae	32	Urticaceae	22	
Polygonaceae	9	Verbanaceae	4	
Portulacaceae	3	Vitaceae	9	

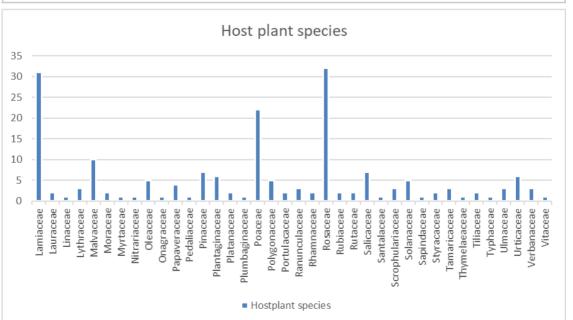


Figure 2. Host plants species of Lygaeoidea species in Turkey.

Table 3. The number of host plant species in Turkey

Hos	t plant Family	Number of Host plant species	Host	plant Family	Number of Host plant species
37	Lamiaceae	31	56	Ranunculaceae	3
38	Lauraceae	2	57	Rhamnaceae	2
39	Linaceae	1	58	Rosaceae	32
40	Lythraceae	3	59	Rubiaceae	2
41	Malvaceae	10	60	Rutaceae	2
42	Moraceae	2	61	Salicaceae	7
43	Myrtaceae	1	62	Santalaceae	1
44	Nitrariaceae	1	63	Scrophulariaceae	3
45	Oleaceae	5	64	Solanaceae	5
46	Onagraceae	1	65	Sapindaceae	1
47	Papaveraceae	4	66	Styracaceae	2
48	Pedaliaceae	1	67	Tamaricaceae	3
49	Pinaceae	7	68	Thymelaeaceae	1
50	Plantaginaceae	6	69	Tiliaceae	2
51	Platanaceae	2	70	Typhaceae	1
52	Plumbaginaceae	1	71	Ulmaceae	3
53	Poaceae	22	72	Urticaceae	6
54	Polygonaceae	5	73	Verbanaceae	3
55	Portulacaceae	2	74	Vitaceae	1

CONCLUSION AND DISCUSSION

In this study, as a result of the literature review between the years 1922-2021, a total of 187 taxa from 38 plant families (67 taxon at genera level, 120 taxon at species level) were identified in association with Lygaeoidea species.

Among these families, Rosaceae has the highest number of host plants, followed by Lamiaceae with 31 taxa number, Poaceae with 22 taxa number and Malvaceae with 10 taxa number (Table 3; Figure 2).

Considering the preference of Lygaeoidea species in these families, 59 species were

associated with Rosaceae, 34 species with Pinaceae, 32 species with Poaceae and Scrophulariaceae, 31 species with Lamiaceae and Oleaceae, 28 species with Tamaricaceae, 26 species with Salicaceae, 22 species with Urticaceae and 15 species with Malyaceae.

When the number of Lygaeoidea species is compared with the plant taxa, the most conspicuous family is the Scrophulariaceae with 32 Lygaeoidea species, related to 3 plant taxa.

It is followed by Tamaricaceae with 28 Lygaeid species related to 3 plant taxa, Oleaceae with 31 Lygaeid species related

to 5 plant taxa, Pinaecae with 34 Lygaeid species related to 7 plant taxa, Urticaceae with 22 Lygaeid species related to 6 plant Atlihan, R., Yardım, N., Özgökçe, S., Kaydan, taxa, and Salicaceae with 26 Lygaeid species related to 7 plant taxa.

Among the plant taxa, Verbascum is taxon most associated with the Lygaeid species, and 37 Lygaeid species have Aysev, N., 1974, Ege Bölgesinde Lygaeidae been identified on the species of this genera. Olea europea with 28 species, Tamarix sp. with 27 species, Pinus sp. with 20 species. and Salix sp., Triticum sp. with 18 species and Urtica sp. follow Beyaz, G., & Tezcan, S., 2002. Studies on the it.

When looking at this list, it can be seen that various species are present on one Cağlar, S., 1992, Beynam ormanı ve çevresiyle or more hosts.

For example, Lygaeus equestris (Linnaeus, 1758) and Spilostethus pandurus in 28, Metopoplax origani (Kolenati, 1845) 24, Çakır, Oxycarenus pallens (Herrich-Schaeffer, 1850) 20. Nysius graminicola graminicola (Kolenati, 1845) and Spilostethus saxatilis (Scopoli, 1763) 16 different hosts were de- Çakır, S., Önder, F., 1990, Turkey Geocorinae tected.

This study (Part-II), some overlooked records that should be included in the part I presented by Kıyak et al (2020) presents as an addition.

With these additional records, the number of families in the Part-I was increased to 39 families with the addition of Asparagaceae, Gençer, N.S, Kovancı, O. B., Kovancı, B., Asphodelaceae and Colchicaceae families, consequently it include 232 taxa with the addition of 10 taxa (4 of at the genera level and 6 at the species level) which were not mentioned in the first study.

As a result of both studies, a total of 409 plant taxa were obtained from 77 plant families associated with Lygaeoidea species in Turkey.

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First record of Sycamore Seed Bug Belonochilus numenius (Say, 1832) (Hemiptera: Heteroptera: Lygaeidae) in Turkish Thrace

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ABSTRACT: In this study, an alien invasive species, Belonochilus numenius (Say, 1832), was recorded in two localities in Edirne province during field studies in the Thrace Region between 2019-2021. These findings are the first record of this species, which entered from Spain to Europe in 2008 and then spread to other European countries, in the Thrace Region and the second record in Turkey. In addition, the latest distribution of this species in Europe and Turkey is shown on the map.

KEYWORDS: Belonochilus numenius, fist record, Lygaeidae, Turkish Thrace.

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INTRODUCTION

the sycamore seed bug, and the natural countries. Belonochilus numenius, which distribution of this Nearctic species is in has been detected in 20 European Canada, the United States, and Mexico countries and Madeira (Aukema, 2020) (Slater, 1964; Ashlock & Slater, 1988).

After this alien and invasive species was identified firstly in Spain in 2008 (Gessé

et al., 2009 the species has spread Belonochilus numenius (Say, 1832) is called rapidly in Central and Southern European has been identified by Çerçi & Oruz (2021) in İzmir Province, which is located in the western Anatolia (Asian part) in Turkey so far.

RESULTS

Belonochilus numenius (Say, 1832)

Material examined: EDİRNE: Uzunköprü-Kırkkavak (41°11'893N; 26°47'591E), In the present study, Belonochilus numenius orientalis L, 1♀.

characteristics of the male ventral views of the species, and its paramera (Fig. 1). Specimens are deposited in the Trakva University zoology collection.

Distrubution in Turkey: İzmir (Selçuk) (Cerci & Oruz, 2021).

General Distrubution: Europe: Albania, Austria, Belgium, Bosnia and Herzegovina, Bulgaria, Croatia, Czech Republic, France, Germany, Greece, Hungary, Italy, Macedonia, Montenegro, Serbia, Sicily, Slovakia, Slovenia,

Spain, Switzerland. North Africa: Madeira. **Asia:** Turkey (Asian part). Extralimital: North America (Canada, Mexico, USA) (Aukema, 2020; Cerci & Oruz, 2021)

25.05.2019, 13; Edirne-Centrum(41°39'597N; was recorded in two localities in Edirne 26°35'789E), 01.07.2021, on Platanus province in Thrace Region. When taking into account the rapid spread of the species, which was also recorded in Bulgaria and The identification was made taking into Greece, it is an expected result that it was consideration the descriptions and the found in the Thrace Region near the borders genitalia of both countries. The species is recorded for indicated by Baena & Torres (2012). In the first time in the Thrace Region, and this the photographs were given dorsal and finding is the second record in Turkey. It seems that this alien invasive species would continue to spread eastward and it would be possible to encounter it in other parts of Anatolia in the future.

> Belonochilus numenius, is an arboreal seed predator, which its adults and nymphs feeding on mature and immature fruits of *Platanus* spp., but it was also occasionally recorded on other deciduous trees in its native area (Wheeler, 1984). In this study, the specimen found in Edirne (Center) was also caught from *Platanus* sp.

Chronological distribution of Belonochilus numenius (Say, 1832) in Palaearctic (Fig.2)

- 2008 France (Matocq, 2008; Baena & Torres, 2012; Dusoulier et al., 2013)
- 2009 Spain (Gessé et al., 2009; Riba et al., 2015)
- 2010 Italy (Küchler & Strauß, 2010)
- 2011 Austria (Rabitsch at al., 2011)
- 2011 Czech Republic (Hradil, 2011; Kment et al., 2013)
- 2012 Hungary (Torma, 2012)
- 2013 Bulgaria (Aukema et al., 2013)
- 2013 Germany (Küchler & Kehl, 2013; Werner et al., 2013; Werner, 2014)
- 2013 Slovakia, (Kment & Cunev, 2013)
- 2015 Madeira (Rabitsch & Heiss, 2015)
- 2016 Bosnia and Herzegovina (Kulijer & Miljevic, 2016)
- 2016 Serbia (Protić & Šeat, 2016)
- 2016 Slovenia (Gogala et al., 2016)
- 2018 Albania (Rabitsch 2018)
- 2018 Greece (Davranoglou & Koutsoukos, 2018)
- 2019 Croatia (Martinović et al., 2019)
- 2019 Macedonia (Srebrova et al., 2019)
- 2020 Belgium (Claerebout et al., 2020)
- 2020 Montenegro (Martinović, 2020)
- 2021 Turkey (Asain part) (Cerçi & Oruz, 2021)
- 2021 Turkey (European part) (this study)

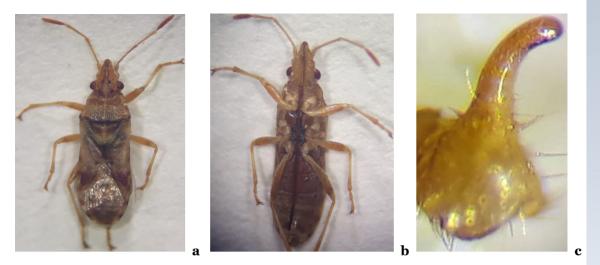


Figure 1. Belonochilus numenius (Say, 1832) a.Dorsal view b. Ventral view c. Paramere

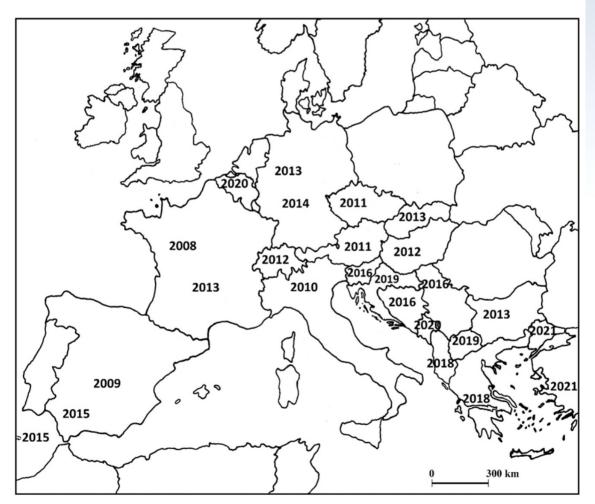


Figure 2. The distribution of Belonochilus numenius (Say, 1832) in Europe.

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Confirmation of the presence of Solenosthedium bilunatum (Lefèbvre, 1827) (Hemiptera: Heteroptera: Scutelleridae) in Albania

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ABSTRACT: Reporting the second record of Solenosthedium bilunatum (Lefèbvre, 1827) in Albania, the presence of the species in the country is confirmed. Information on the known distribution of S. bilunatum in Europe is summarized.

KEYWORDS: Solenosthedium bilunatum, distribution, Albania.

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So far, Solenosthedium (Lefèbvre, 1827) (Hemiptera: Heteroptera: from Albania so far, which was found in Scutelleridae), a Mediterranean scutellerid Kokome (Sarandë) on 31.10.1970, was belonging to the subfamily Elvisurinae, reported by Misja (1973). has been reported from the following European countries: Albania, Croatia (mainland, Korčula), France Cyprus, (mainland, Corsica), Greece, Italy (mainland, Sardinia, Sicily, Ustica), Malta, Portugal and Spain (mainland, Ibiza, Mallorca) (Misja, 1973; Josifov, 1986; Matocq & Pluot-Sigwalt, 2002; Göllner-Scheiding, 2006; Gogala, 2008; Aukema et al., 2013; Dusoulier et al., 2016; Škorput et al., 2019; van der Heyden, 2020).

bilunatum The only known record of S. bilunatum

Now, the presence of S. bilunatum in Albania can be confirmed: On 29.09.2021, a nymph was photographed by Théalie Dhellemmes on Paliurus spina-christi Mill. (Rhamnaceae) near the city of Sarandë, located at the Ionian Sea (Fig. 1). The photograph was uploaded to the online database iNaturalist (under the pseudonym thealie) (2021).

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Figure 1. Nymph of *Solenosthedium bilunatum* (Lefèbvre, 1827), near Sarandë, Albania, 29.09.2021. (Photo: Théalie Dhellemmes).



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An Additional Locality Record for the Rare Distributed *Pasira* marinadolina P.V. Putshkov & Moulet, 2004 (Hemiptera: Heteroptera: Reduviidae) in Turkey with Description of Macropterous Female

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ABSTRACT: In this study, a macropter female specimen of *Pasira marinadolina* P.V. Putshkov & Moulet, 2004 was determined in Edirne province in 2021. This finding is the second locality record of this species in Turkey. In this study, the description of the macropter form is given and the differences between it and the related species *Pasira basiptera* Stål, 1859 are discussed.

KEYWORDS: Pasira marinadolina, macropterous female, description, Turkey.

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INTRODUCTION

There are 4 species of the genus *Pasira* Stål, 1859 (Heteroptera: Reduviidae: Reduviinae) in the Palearctic Region, and 3 of them are distributed in the Western Palearctic (Aukema, 2020). Of these species, *Pasira perpusilla* (Walker, 1873) is distributed in the southwestern part of

China in the Eastern Palearctic. *Pasira basiptera* Stål, 1859, one of the species distributed in the Western Palearctic, is the most widely distributed species of the genus, and its distribution is known from southern Europe and especially in the Balkans, in North Africa and in the Middle East, Arabian Peninsula, Caucasus

known until now from Ukraine and the bristles, veins prominent. European part of Turkey in Europe and Connexivum unicolar, yellowish brown. (Aukema, 2020).

Pasira Stål, 1859 is represented by two yellow. Size 6,1mm. (Fig. 1). species in Turkey. Of these, Pasira basiptera has only been given from Anatolia so far (Dursun & Salur, 2013).

province in Turkey.

Pasira marinadolina P.V. Putshkov & Moulet, 2004

Material: Edirne: Center (Trakya University-Balkan Campus), 20.05.2021, 1 macropterous female.

Distribution in Turkey: İstanbul-Esenyurt (Çerçi & Koçak, 2016), Edirne (the present study)

General Distribution: Europe: Turkey 2020).

Head black, brownish in distal, with, the only sample we have is female. Apart golden bristles equal to the length of the from male genitalia, diameter of the first segment of the is that P. marinadolina has two oblique sulcus in dorsal; anterior part in front of the pronotum. This the sulcus about twice as long as its given by P.V. Puthkov & Moulet (2009) for posterior part; first two segments of the pronotum of micropter form is also yellowish brown. posterior margin thinly lighter - reddish brown, only the proximal corners and brown; anterior part about 1.2 times as margins are lighter reddish-brown (Fig. long as posterior part; anterior lobe of 1). However the pronotum in the macropter the pronotum with two oblique lateral form of P. basiptera is chocalat brown, impressions, as in the micropter form of strangled in its middle, this hollow disc this species Scutellum brownish black, with a strong transverse groove seperated with rare and short bristles; the carina- two sub-equal lobes. Pronotum bulging in shaped lateral edges joined distally and the anterior, with a medio-longitudinal elongated and formed a "Y"-shaped short furrow generally not reaching the structure, which is much better seen in posterior edge and delimiting two erased

in western Asia and in central Asia. the lateral view. Corium in proximal (up Pasira lewisi (Miller, 1951) is distributed to the distal part of clavus) reddish only in the Canary Islands in the Palearctic yellow (lighter in distal of clavus) the rest Region. The presence of Pasira marinadolina (including membrane) brown, lateral P.V. Putshkov & Moulet, 2004 has been margins and dorsal with short and golden

from Azerbaijan and Armenia in Asia Legs, reddish yellow with short golden bristles, tibiae slightly darker, tarsi pale

Comments and remarks: This species was previously given only based on on brachypter specimens (P.V. Putshkov & Pasira marinadolina P.V. Putshkov & Moulet, 2004, 2009; Çerçi & Koçak, Moulet, 2004 was recorded for the first 2016). The record given by Cerci & time by Çerçi & Koçak (2016) from Esen- Koçak (2016) from İstanbul in the yurt in the European side of Istanbul European part of Turkey is based on 6 brachvpyter specimens of marinadolina. While the authors listed the differences of this species from Pasira basiptera, they reported that P. marinadolina was always micropterous unlike basiptera. However, as later presented its photographs on the Doğal Hayat website by Barış Çerçi, and our study shows that P. marinadolina also has macropter forms.

Remarks: P.V. Puthkov & Moulet (2009) stated that the only known micropter (European part), Ukraine. Asia: Azerbaijan, form of *P. marinadolina* is very similar to Armenia, Turkey (Asian part) (Aukema, P. basiptera, except for the male genitalia. Unfortunately, we were not able to compare Description of macropterous female: the species in terms of male genitalia, as another difference and very deep transverse lateral impressions on anterior lobe of diagnostic character Pronotum seen in the macropter form. Additionally, posterior corners and the pronotum is completely blackish-





Figure I. Pasira marinadolina P.V. Putshkov & Moulet, 2004. a. Habitus (dorsal view) b. Head, pronotum and scutellum (dorsal view)

bumps in the posterior; posterior margin **Ecology**: P. marinadolina occurs on convex and narrowly thinned (See P.V. stony slopes covered by sparse vegetation. Putshkov & Molulet, 2009 p. 514, Fig. 95 Adults are mostly found under stones -a).

Other differences mentioned for the species of genus Pasira in the key given by P.V. Putshkov & Moulet (2009) are on the colorations of the connexivum. The connexivum in P. basiptera is bicolored and reddish or brownish with black spots, rarely completely yellow/yellowish. The pronotum in P. marinadolina is one colored and is given as yellow or yellowish by the authors (P.V. Putshkov & Moule, 2009). In our specimen, the connexivum is yellowish brown as one colored. Cerci & Koçak (2016) stated that this character is not valuable because the connexivum coloration of microptereous specimens of Aukema, B., 2020, Catalogue of Palaearctic P. marinadolina in their study is bicolored as in P. basiptera.

from September to June, but they also live in plant debris, under bushes and overwinter as an adult (Puthkov Moulet, 2009). Puthkov & Moulet (2009) reported that they collected specimens of P. marinadolina at the entrance of a fox hole. In this study, a single specimen was caught inside the building of Biology Department in the Trakya University Campus. Since it was cold and rainy in May and June of 2021, this individual was probably inside the building to protect from the cold.

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The sensilla on head, antenna and mouth parts in Aelia rostrata Boh. (Hemiptera, Pentatomidae): A scanning electron microscopical study

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ABSTRACT: The sensilla who act as thermohygroreceptor, chemoreceptor, or mechanoreceptor are the main sensory structures of insects. Insects carry different types, numbers and distributions of sensilla in body parts such as head, antenna, mouth parts, and leg segments. In this study, the sensilla types on the head, antenna and mouth parts of Aelia rostrata Boh. (Hemiptera, Pentatomidae) were invastigated with using scanning electron microscope. According to the results of the study, 4 major types of sensilla with different diameters and cone-shaped protrusions have been identified: sensilla basiconica, sensilla trichodea, sensilla peg, and sensilla campaniformia. These sensilla show different distributions in the examined structures of the insect. The results obtained from the study were also compared with other species in the literature, and similarities and differences were revealed.

KEYWORDS: Chemoreceptor, Heteroptera, insect, mechanoreceptor, morphology.

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INTRODUCTION

Order Hemiptera is an extensive group that includes plant pests. The species belong to order Hemiptera have piercingsucking mouth parts which gives insects bug is mainly because it is a wheat pest. an advantage in feeding on plant sap In

Aelia rostrata Boh. (Hemiptera, Pentatomidae) which is known as wheat stink bug is a significant pest in Turkey.

The reason why it is called a wheat stink addition, rye, barley, oats and some (Hao et al., 2016; Kanturski et al., 2017). other graminaceous plants are also infested by this insect.

A. rostrata is a widely distributed and thermohygrosensory (Fernandes et migratory species is dos, 1981; Lodos et al., 1984; Önder et porous (Nowinska & Brozek, 2017). al., 1995; Özgen et al., 2005; Gözüaçık et al., 2011; Khaghaninia et al., 2013; Tarla, 2017; Bolu, 2020).

sensilla on their mouth parts and antenna 2017; Taszakowski et al., 2019; Amutkan which enter into crucial role in some Mutlu et al., 2021). functions such as feeding or mating [(e.g. the sensilla on the mouth parts scan the surfaces of food (Chapman, 1998; Brozek & Zettel, 2014; Parveen et al., 2015) and antenna which is the main sensory of insects, detecting chemicals in the air (Carey & Carlson, 2011; Rani et al., 2021)] (Isidoro et al., 2001; Fu et al., 2012; Cao & Huang, 2016; Seada & Hamza, 2018; Faucheux et al., 2020).

distance (Brozek, 2013). Together with All male and female individuals were the different types of sensilla on the adults. antenna, they function as chemoreceptors, thermohygrorecertors, and mechanoreceptors (Akent'eva, 2008; Fu et al., 2012; Brozek & Bourgoin, 2013; Freitas et al, 2020; Giglio et al., 2021; Zhang et al., 2021).

Many different types of classification can be made in sensilla in insects according to different criteria. For example, according to their perception ways, they are divided into 2 main groups as mechanoreceptors. All studies were performed at Gazi University, and chemoreceptors which are more Faculty of Science, Prof. Dr. common (Brozek & Chlond, 2010; Li et Suludere Electron Microscope Center. al., 2016). Sensilla can also be classified according to their morphological features, such as basiconic, placoid, trichoid, long RESULTS AND DISCUSSION hair-like, plate-like or coeloconic, etc. (Slifer 1970; Altner & Prillinger, 1980; Hallberg & Hansson, 1999; Shields 2010; Nowinska & Brozek, 2017).

located in Adıya- al., 2008; Nowinska & Brozek, 2017; Li et man, Batman, Diyarbakır, Gaziantep, al., 2018). Another classification is based Mardin, Sanlıurfa, and Sırnak provinces on whether sensilla carry pores or not. in Turkey. This species has also been According to this, they are collected in 3 found in Iran and Iraq (Brown, 1965; Lo- groups as aporous, uniporous and multi-

The morphological structure of the sensilla on the mouth parts and antenna varies among Hemiptera species (Brozek & Insects have some structures called Bourgoin, 2013; Nowinska & Brozek,

> The main goal of this study is to disclose the morphological features of the cuticular structures of sensilla on the antenna and the mouth parts in wheat stink bug A. rostrata.

MATERIALS AND METHODS

The male and female individuals of Aelia rostrata (Figures 1A-B) were collected in Sinanlı and Oltan (Ayaş) and Çağa Village So, the insect recognizes plants at a (Güdül) in Ankara province in July, 2018.

> First, the external surface of the integument of insects was cleaned. Then, the specimens were dried in the air and adhesive o the stubs.

> Subsequently, the stubs were coated with gold with Polaron SC502 sputter coater, observed in JEOL JSM 6060 LV SEM, and photographed (at 10-15 kV accelerating voltage).

Insects have sensory organs called sensilla, which enable the detection of chemical substances in their environment in various parts of their bodies (Cao & Huang, 2016; Besides, insect sensilla can be classified Amutkan Mutlu et al., 2021). In this according to their sensory modality, such study, sensilla types found on the head, as olfactory, gustatory, mechanosensory, antennae and mouth parts of male and female *A. rostrata* individuals were types. In this context, the mean values of investigated with SEM. The diagrams of the sensilla trichodea and the sensilla different types of sensilla on mouth parts, basiconica lengths are given in Table 1 antenna, and head of *A. rostrata* are and Table 2. In addition to these types of shown in Figure 2. As a result of the sensilla, sensilla peg (Sp) and sensilla study, it was observed that the sensilla campaniformia (Sca) were also detected trichodea (St) and the sensilla basiconica as a result of our study. (Sb) were more common than other sensilla

Table 1. Mean values of St lengths in both males and females (µm)

			Mouth par	rts		
	Labium 1	Labium 2	Labium 3	Labium 4	Labrum	
Male	45,1 ± 19,6	55,0 ± 15,6	41,3 ± 21,8	-	46,8 ± 5,4	
Female	61,0 ± 23,2	40,3 ± 10,4	53,4 ± 21,1	-	-	
			Antenna	ı		
	Scape	Pedicel 1	Pedicel 2	Flagellum 1	Flagellum 2	
Male	-	33,0 ± 5,6	39,3 ± 13,8	21,1 ± 9,5	24,5 ± 8,4	
Female	-	29,9 ± 6,6	-	$28,3 \pm 6,0$	33,7 ± 3,0	
	Head					
	1	2	3	4		
Male	78,9 ± 20,4	82,7 ± 26,6	$71,0 \pm 22,7$	-	-	
Female	37,8 ± 24,6	83,9 ± 22,1	52,5 ± 20,6	-	-	

Table 2. Mean values of Sb lengths in both males and females (μm)

			Мо	uth p	arts	
	Labium 1	Labium 2	Labium	1 3	Labium 4	Labrum
Male	-	42,1 ± 6,2	-		2,9 ± 0,6 (sho 12,3 ± 2,9 (ta	,
Female	-	-	-		3,8 ± 0,8 (sho 14,3 ± 1,6 (ta	
			A	nteni	na	
	Scape	Pedicel 1	Pedice	el 2	Flagellum	1 Flagellum 2
Male	-	-	-	- 50,4 ± 7,8		37,7 ± 12,4
Female	-	-	51 ± 1	$51 \pm 11,1$ $11,2 \pm 2,3$		10,3 ± 1,9
	Head					
	1	2	2		3	4
Male	-	-	-		-	-
Female	-	-	-		-	-

Sensilla types on the mouth parts

The mouth parts are generally consisted of the labrum (Lm), the labium (Lb), and a labial groove in hemipteran insects But the second segment is thinner and (Figures 3A-B). Although the general longer than the third segment. In both structure of these parts is very similar, sexes of A. rostrata, the second segment differences can be observed in their of the labium has sensilla trichodea, detailed structure, number of segments, sensilla basiconica and sensilla campanisensilla types and distributions among formia (Figures 4A-F). different species.

The mouth parts of A. rostrata males and campaniformia can be also observed on females have a labrum with proximal and the third segment of labium in both distal sides, four segmented labium, sexes. labial groove, and stylet fascicle which is the defining characteristic of hemipterans species (Wang et al., 2020a; Amutkan Mutlu et al., 2021).

Piezodorus hybneri (Gmelin, 1790) (Hemiptera, Pentatomidae), Perillus bioculatus (F.)(Hemiptera, Pentatomidae), Eocanthecona furcellata (Wolff) (Hemiptera, Pentatomidae), Dolycoris indicus Stål, 1876 (Hemiptera, There are sensilla trichodea and sensilla Pentatomidae), Macrocheraia grandis (Gray, quadriguttata Bergroth, 1894 (Hemiptera, F). Largidae), Physopelta gutta (Burmeister, 1834) (Hemiptera, Largidae), Physopelta cincticollis Stål, 1863 (Hemiptera, Largidae), and Cheilocapsus nigrescens (Liu and Wang) (Hemiptera, Miridae), the labium have four segmented as in A. rostrata, but The labrum has two regions according to Eurygaster testudinaria has segmented labium in its mouth parts regions. (Parveen et al., 2015; Wang et al., 2019, 2020a; Amutkan Mutlu et al., 2021).

segmented in both male and female A. rostrata. The first segment is concave and the labrum (Lm) runs parallel to the first segment of the labium (Figures 3A-B).

Therefore, looking at the insect from the ventral side, the labrum is in the middle and the first segment of the labium (Lb1) lie on either side of it. The first segment of labium has many sensilla in different sizes such as sensilla trichodea and Sensilla types on the antenna sensilla campaniformia (Sca) in both male and female insects.

But, sensilla campaniformia is more common segment has sensilla of different types, in females than males (Figures 3C-F).

The second and third segments of the labium (Lb2 and Lb3) are seen verv similar to each other.

sensilla The trichodea and

However, there are differences in the morphological structures of the sensilla trichodea and sensilla campaniformia in males and females (Figures 5A-F).

In the fourth segment of the labium (Lb4), sensilla density is higher than in other segments.

basiconica on the fourth segment of the 1832) (Hemiptera, Largidae), Physopelta labium of females and males (Figures 6A-

> The labrum (Lm) is the part of the mouth which attaches to the anterior region of the head and extends to first segment of labium (Figures 3A-B).

three- the surface patterns: proximal and distal

The proximal region is the part that is close to the region where the labrum at-The labium (Lb) is long, thin, and four taches to the head and has smooth surface.

> Very few sensilla trichodea are seen on this area in both sexes (Figures 7A-B).

> The distal region is the part near the free end with transverse crests and has no sensilla in both males and females (Figures 7C-D).

In A. rostrata, the antenna is divided into 5 segments in both sexes and each density and distribution (Figures 8A-B).

On the surface of the scape, there are (Figures 11A-D). numerous one or two apex cone-shaped protrusions.

Also in this region there are only a few peg, and sensilla campaniformia types. long sensilla peg (Sp) in males and females (Figures 8C-H).

different varieties of sensilla campaniformia D). in male individuals.

Although the sensilla in females are very similar to those in males, they have only 1 type of sensilla campaniformia (Figures 9A-F).

The density of sensilla on the pedicel 2 is much higher than the scape and pedicel 1.

Sensilla basiconica, sensilla trichodea, and two types of sensilla campaniformia were observed in both females and males. but sensilla basiconica is the predominant sensilla type in this segment (Figures 10A-B).

The highest number of sensilla is on the flagellum 1 and 2. In addition, sensilla variety is more in these segments. Different sizes of sensilla basiconica, sensilla peg female individuals. In addition, in males on the surface (Figures 14C-F). there is also sensilla campaniformia

Flagellum 2 of male insects has sensilla basiconica, sensilla trichodea, sensilla

In females, only sensilla basiconica, sensilla trichodea, and sensilla peg are Sensilla trichodea is found on pedicel 1, found, and the bases of some sensilla albeit rarely in males. There are also two basiconica appear swollen (Figures 12A-

Sensilla types on the head

As a result of the SEM analysis, it was determined that there are many different types of sensilla on the head regions of A. rostrata males and females.

The head can be divided into 4 regions, as in the figure, according to the sensilla on its surface (Figures 13A-B).

Sensilla campaniformia and different types of sensilla trichodea are found in the first and the second regions of the head in both males and females (Figures 13C-F).

In both sexes, only sensilla trichodea was seen on the third region of the head (Figures 14A-B).

and two types of sensilla trichodea were In the fourth part, there are numerous detected in the flagellum 1 in male and one or two pointed cone-shaped protrusions





Figure 1. Stereomicroscope photograph of A. rostrata. A. Female individual, B. Male individual (Scale bar: 1 mm)

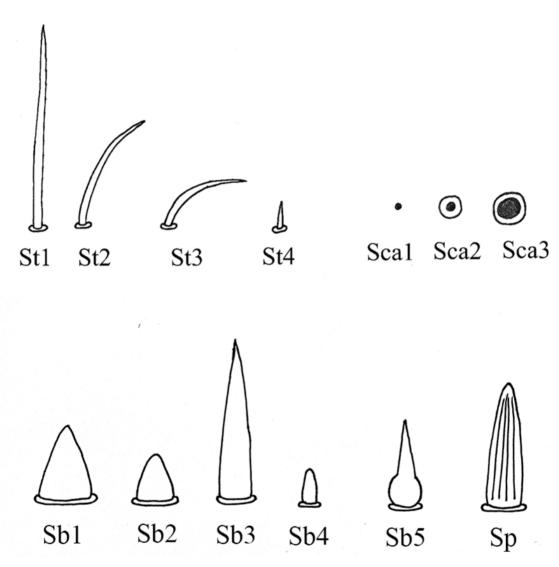


Figure 2. The schematic illustration of different types of sensilla on antenna, mouth parts and head of *A. rostrata*. St1: sensilla trichodea 1, St2: sensilla trichodea 2, St3: sensilla trichodea 3, St4: sensilla trichodea 4, Sb1: sensilla basiconica 1, Sb2: sensilla basiconica 2, Sb3: sensilla basiconica 3, Sb4: sensilla basiconica 4, Sb5: sensilla basiconica 5, Sp: sensilla peg, Sca1: sensilla campaniformia 1, Sca2: sensilla campaniformia 2, Sca3: sensilla campaniformia 3

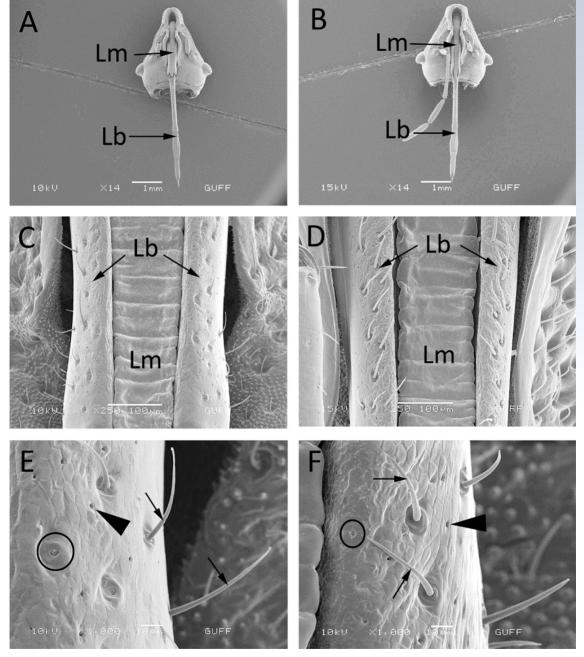


Figure 3. A-B. General view of the mouth parts, A: Female, B: Male. C-F. The first segment of the labium, C-E: Female, D-F: Male. Lb: labrum, Lm: labium, arrow: sensilla trichodea 2, arrowhead: sensilla campaniformia 1, encircled: sensilla campaniformia 2.

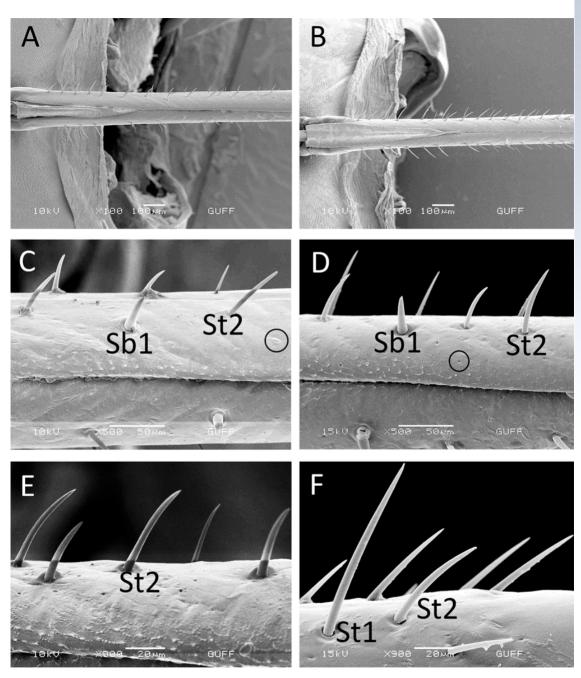


Figure 4. The second segment of the labium. A-C-E: Female, B-D-F: Male. Sb1: sensilla basiconica 1, St1: sensilla trichodea 1, St2: sensilla trichodea 2, encircled: sensilla campaniformia 1.

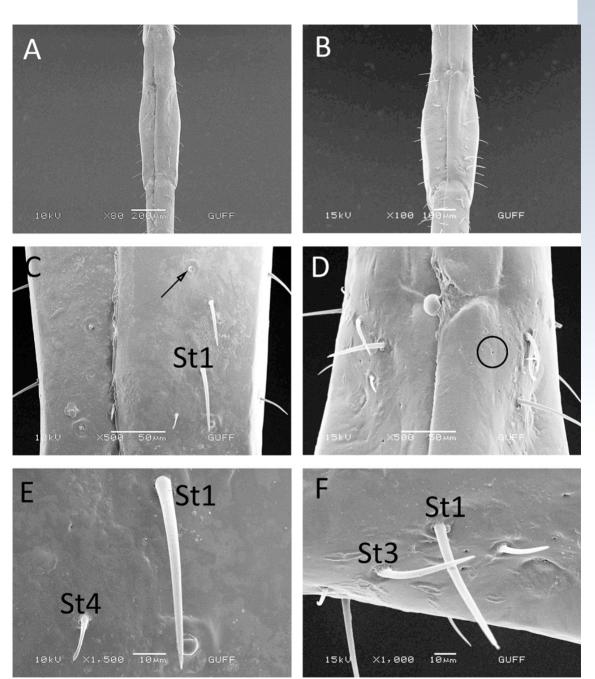


Figure 5. The third segment of the labium. A-C-E: Female, B-D-F: Male. St1: sensilla trichodea 1, St3: sensilla trichodea 3, St4: sensilla trichodea 4, encircled: sensilla campaniformia 1, arrow: sensilla campaniformia 2.

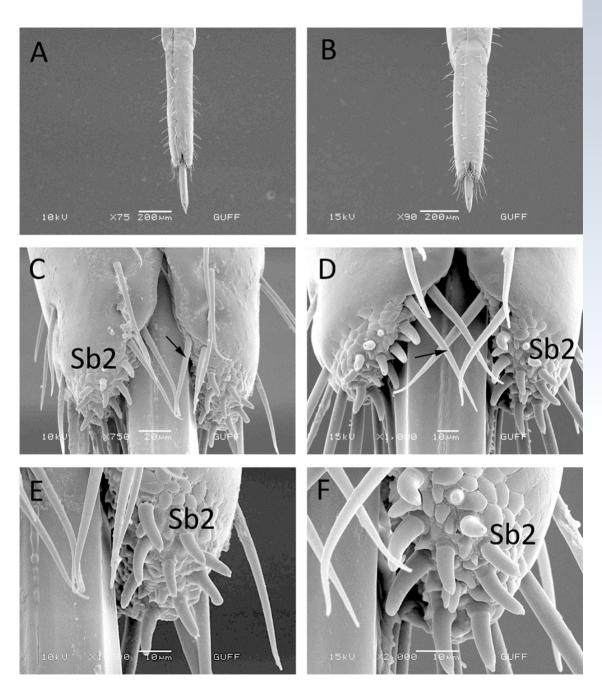


Figure 6. The fourth segment of the labium. A-C-E: Female, B-D-F: Male. Arrow: sensilla trichodea 1, Sb2: sensilla basiconica 2.

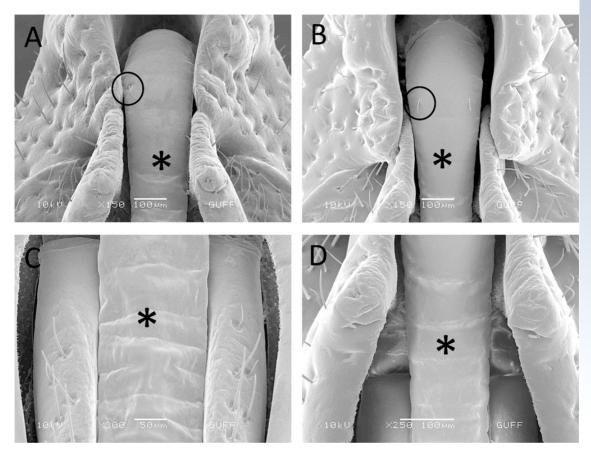


Figure 7. A-B. The proximal region of the labrum, C-D. The distal region of the labrum. A-C: Female, B-D: Male. Encircled: sensilla trichodea 3, asterisk: labrum.

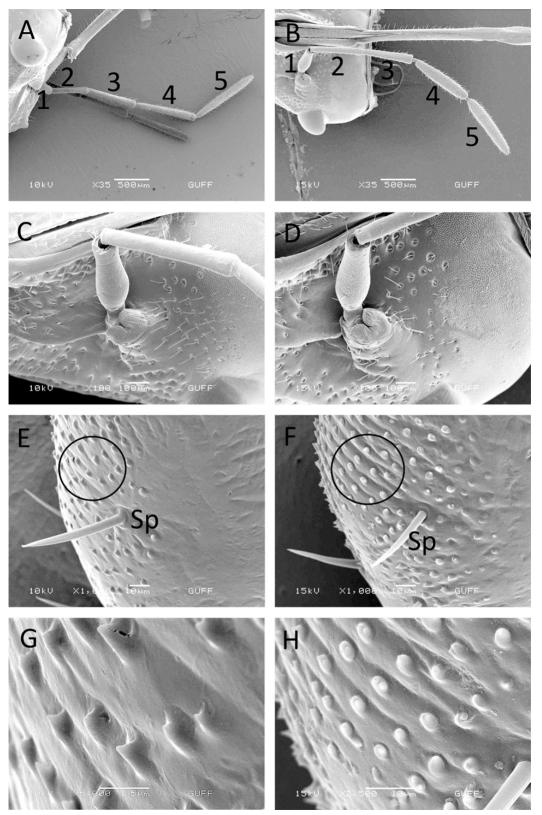


Figure 8. A-B. General view of the five segment of the antenna. C-H. The scape of the antenna. A-C -E-G: Female, B-D-F-H: Male. Sp: sensilla peg, encircled: cone-shaped protrusions.

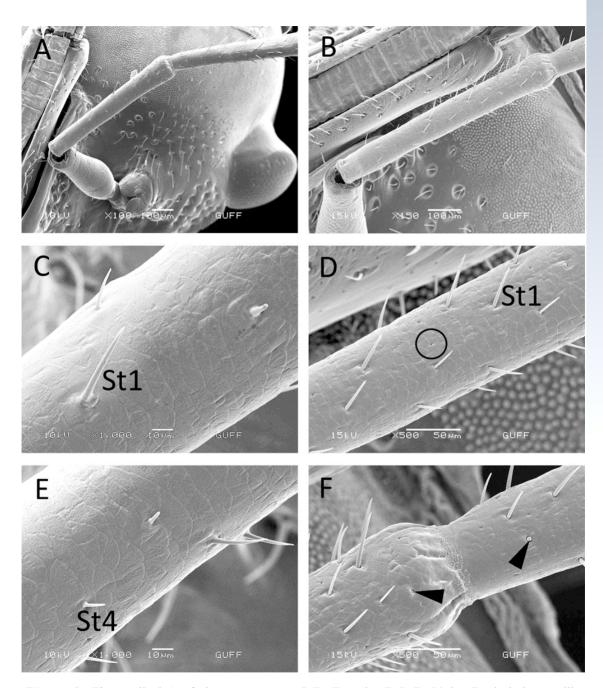


Figure 9. The pedicel 1 of the antenna. A-C-E: Female, B-D-F: Male. Encircled: sensilla campaniformia 1, arrowhead: sensilla campaniformia 2, St1: sensilla trichodea 1, St4: sensilla trichodea 4.

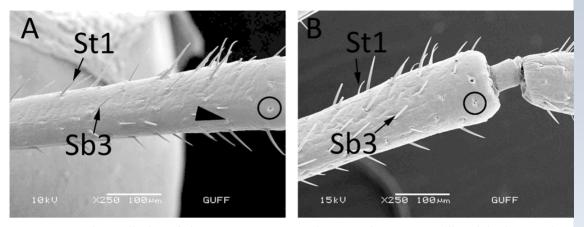


Figure 10. The pedicel 2 of the antenna. A: Female, B: Male. St1: sensilla trichodea 1, Sb3: sensilla basiconica 3, arrowhead: sensilla campaniformia 1, encircled: sensilla campaniformia 2.

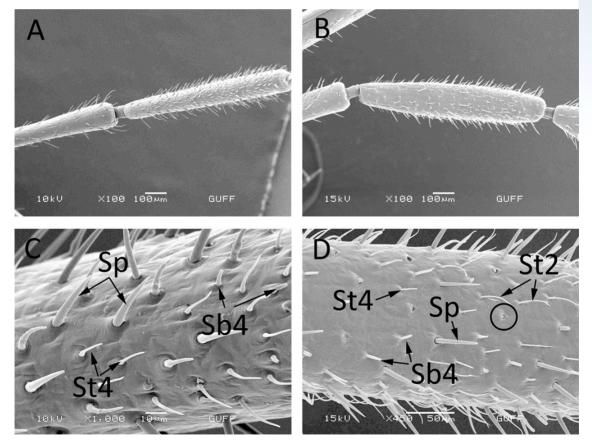


Figure 11. The flagellum 1 of the antenna. A-C: Female, B-D: Male. St2: sensilla trichodea 2, St4: sensilla trichodea 4, Sp: sensilla peg, Sb4: sensilla basiconica 4.

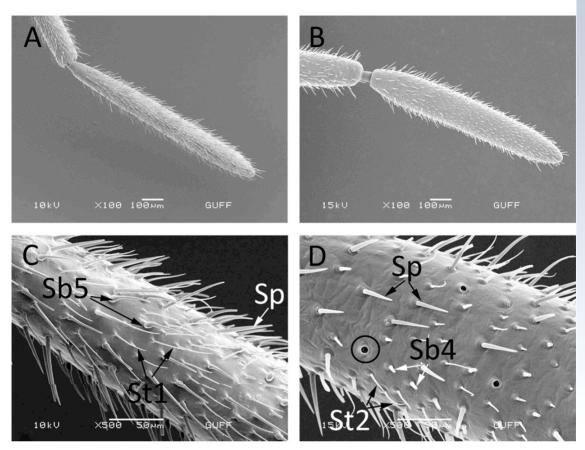


Figure 12. The flagellum 2 of the antenna. A-C: Female, B-D: Male. St1: sensilla trichodea 1, St2: sensilla trichodea 2, Sb4: sensilla basiconica 4, Sb5: sensilla basiconica 5, Sp: sensilla peg, encircled: sensilla campaniformia 3.

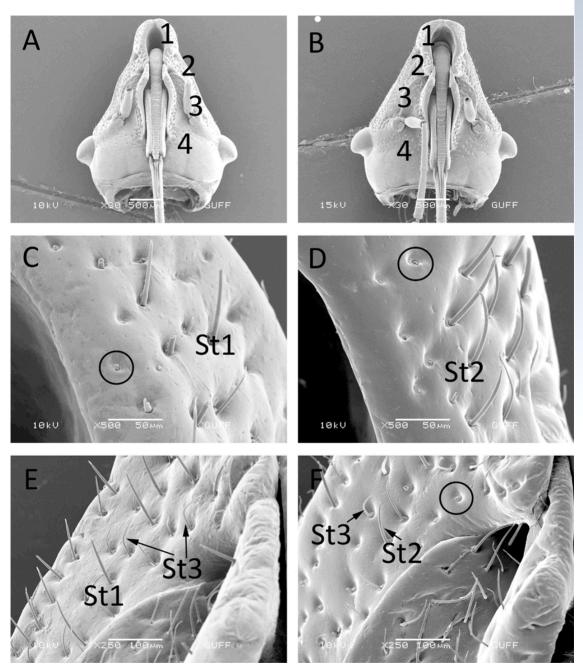


Figure 13. A-B. General view of 4 regions of the head. C-D. The first region of the head. E-F. The second region of the head. A-C-E: Female, B-D-F: Male. St1: sensilla trichodea 1, St2: sensilla trichodea 2, St3: sensilla trichodea 3, encircled: sensilla campaniformia 2.

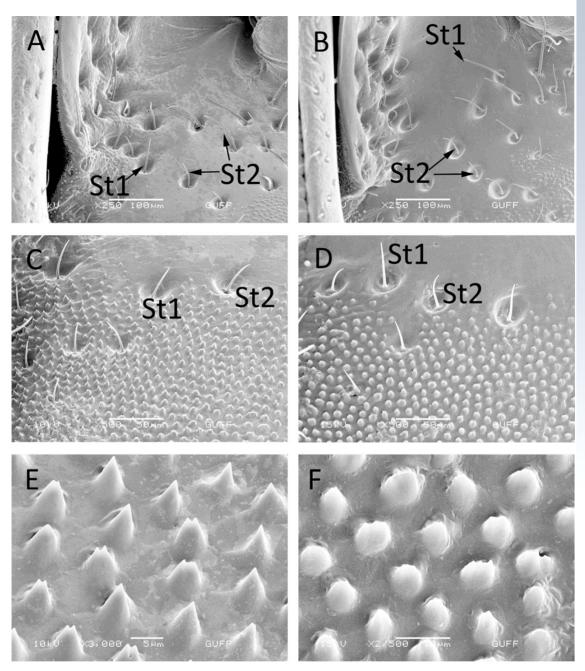


Figure 14. A-B. The third region of the head. C-F. The fourth region of the head. E and F shows the one or two pointed cone-shaped protrusions. A-C-E: Female, B-D-F: Male. St1: sensilla trichodea 1, St2: sensilla trichodea 2.

Sensilla and distributions in mechanoreceptor types insects are thought to be related to the ensures the discovery of nutrients (Liang insects' diet. Because, each sensilla has et al., 2013; Gullan & Cranstone, 2014; different sensory functions. For example, Wang et al., 2019; Amutkan Mutlu et al., sensilla basiconica in the mouthparts is 2021). Comparison of sensilla types of responsible for the movement of these some species in the literature is given in parts, while sensilla trichodea acts as a the Table 3. It is seen in the table that

and in and thus to the structure of their mouth- subject.

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sensilla types can vary in different parts. The fact that it has so many species of insects. We can say that sensilla different types of sensilla shows that it varieties, numbers and distributions give can receive various chemical stimuli in A. us taxonomic and phylogenetic data, rostrata. We hope that this study on the especially thanks to their morphological sensilla of A. rostrata will contribute to contributions to the nutrition of insects other studies and literature on this

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	Mouth parts	n				Antenna						
Species	Labium 1	Labium 2	Labium 3	Lebium 4	Labrum	Scape	Pedicel1	Pedicel2	Flagellum 1	Flagellum2	Head	Reference
Aeda rostrata	5t, 5ca	St, Sb, Sca	St. 5ca	St, 50	វ	Sp. cone- shaped protrusions	St, Sca	St, Sb, Sca	St, Sb, Sp	St, Sb, Sca, Sp	Sca, St, cone- shaped protrusions	
Dolycon's indicus	,							,	St, Sb, Sca, Sch**	3ch**		Ahmad et al., 2016
Plactia crossota	,								St, Sb, Sca			Ahmad et al., 2016
Eucanthecona percellata	,					-	,		St, Sb, Sca, Scorre, Spirms	Sco***.	-	Abmad et al., 2016
Peritus bioculatus	5p, Sstc*, St, Sb, Sca	t, 5b, 5ca					,		St, Sb, Sca, Sco	500		Farven et al., 2015; Abmad et al., 2016
Tricsokus japonicus						ĸ	Sb, St		St, Sch, Spa sickle-shape peg sensilla	St, Sch, Spap****, Sca, sickle-shaped, groowed- peg sensitia		Yang et al., 2016
Trissokus plantiae	,					ĸ	Sp. St		St, Sch, Spe -shaped, gro sensilia	St, Sch, Spap, Sca, sickle -shaped, gromed-peg sensilia		Yang et al., 2016
Stephanitis nashi						5t, 5co	St, 5b, Sca		ម	St, grooved- peg sencita, SCo		Wang et al., 2020b
Hazyomorpha hatys	,					Sb, 5co, 5ch	Sb, Sco, Sch	5b, 5co, 5ch	5b, 5t, 5co, 5ch	5b, 5t, 5co, 5ch		Ibrahim et al., 2019
Euschistus heros	,						8	Sb	St, Sb	St, 5b		Silva et al., 2010
Edessa meditabunda	,						Sb, Sch	Sb, 5ch	St, Sb	St, Sb		Sìta et al., 2010
Piezodorus guildīnii							Sb, Sch	Sb, Sch	St, Sb	St, Sb		Silva et al., 2010
Eocarthecona parcellata	Sp, St, Sstc, Sb, Sch	, Sb, Sch				Sch, Sb	St, Sch, Sb	St, Sch, Sb, Sco	5t, 5ch, 5b, 5co	St, Sch, Sb, Sco	,	Farven et al., 2015; Zhao et al., 2021
Dolycoris indicus	5p, 5t, 5b, 5ca, 5cb	Sca, Sch										Parven et al., 2015;

Table 3. Comparison of sensilla types of A. rostrata and some species belong to the order Hemiptera in the literature.

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Table 3. Continued	pg.											
Plautia crossota	Sp. St. Sb, Sca, Sch	Sch			-							Farreen et al., 2015;
Piezodorus hybemi	Sp, St, Sb, Sca, Sch	Sch			-							Parreen et al., 2015;
Eurygaster testudinaria	no sensilla	5b, 5t, 5ca, 5p	5b, St, Sca		dome-shaped protrusions, plate-shaped structures	Sb, St, Sca, Sp	5b, 5t, 5ca, 5p	5b, 5t, 5ca, 5p	5b, 5t, 5ca, 5p	5b, 5t, 5ca, 5p	St, Sca,	Amutkan Mutiu et al., 2021
Arma chinensis				,		Sb	SS SS	Sb, Sch	5t, 5b, Sch	5t, 5b, 5ch, 5ca, 5co		Zhang et al., 2014

*: sensilla styloconica, **: sensilla chaetica, ***: sensilla coeloconica, ***: sensilla placoidea, ****: sensilla papillary



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Contributions to the Nabidae (Hemiptera: Heteroptera) Fauna in the Thrace Region, Turkey

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ABSTRACT: The present study was conducted in 73 localities between May and September of 2013 and 2014 in Edirne Province and in two localities in in 2016 in Kırklareli Province. As a result, 6 species belonging to 3 genera from Nabinae and Prostemmatinae subfamilies of Nabidae family were determined. Among these species, Nabis flavomarginatus Scholtz, 1847 and Nabis ferus (Linnaeus, 1758) are first records for the Heteroptera fauna of the Thrace Region and additional locality records for 4 species known from the region are given.

KEYWORDS: Nabidae, fauna, taxonomy, first record, Turkish Thrace, Turkey.

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INTRODUCTION

All species of the Nabidae family are predators and feed on eggs, larvae and adults of insects and other small Arthropods.

Most species are polyphagous, species of the subfamily Prostemmatinae pests (Kerzhner, 1996). particularly feed on Lygaeidae, another family of Heteroptera. All Prostemmatinae and some Nabinae species live in soil and are found among debris or under stones. Some Nabinae species are found on grasses and some on trees.

Most species spend the winter as adults some as eggs. Some belonging to the subfamily Nabinae (Nabis ferus, N. pseudoferus, N. punctatus, N. palifer, N. capsiformis and Himacerus apterus) are more or less important in but the control of agricultural and forest

Nabidae family is a small family and is represented with 500 species belonging to 21 genera in the world, 122 species belonging to 3 subfamilies and 10 genera in the Palearctic Region and 26 species belonging to 2 subfamilies and 5 genera in Turkey (Kerzhner, 1996; Önder et al., MATERIAL AND METHOD 2006; Aukema et al., 2013).

Although the studies on the Nabidae family native researchers (Horváth, 1883, 1901, Edirne Province between 1981, 1983, 1984, Kıyak, 1990) gave was added the study data (Table 1). local records limited to a few species. while Erbay (1986) mentioned from species in his master's thesis on Nabidae family collected from different localities from Turkey, and Dursun (2011) from 9 species in his study on Kelkit Valley Heteroptera species.

1984; Dursun & Fent, 2015). family is represented by 42 species in Europe, 15 species in Greece and 17 species identification of the species. in Bulgaria (Aukema, 2013).

The research material belonging to the Nabidae were collected from family in Turkey started in the old years, different localities in terms of habitat they are quite limited. The foreign and characteristics, which were determined in 1905, 1918; Reuter, 1890; Puton, 1892; September in 2013-2014years, especially Puton & Noualhier, 1895; Kiritshenko, during the spring-summer period when 1918, 1924; Fahringer, 1922; Gadeau De the species are active. In addition, the Kerville, 1939; Hoberlandt, 1956; Linnayuori, material obtained from two locations 1965, Tuatay et al., 1972; Önder et al., (loc.74, loc.75) from Kırklareli Province

The research material was collected with the help of a sweep net from short herbaceous plants and with an aspirator from the soil. In addition, male pygophore and paramer structures are highly useful morphological characters in Nabidae family species, as in most Heteroptera The records of the Nabidae family from suborder species. For this reason, after the Thrace Region are very limited and so the male specimens were kept in hot far 10 species belonging to 4 genera have water for about 5 minutes, the pygophore been recorded (Horváth, 1918; Fahringer, and its paramers were removed under the 1922; Hoberlandt, 1956; Önder et al., microscope with the help of a fine-tipped Nabidae forceps and a preparation needle, and

these structures were also used in the

Table 1. The localities, coordinates, altitudes and the dates of the field studies in Edirne Province

Loc No	Locality	Coordinate	Altitude	Date
1	Havsa-Arpaç	41°41'08N 26°52'33E	113m	30.08.2013
2	Havsa-Necatiye	41°30'18N 26°53'54E	57m	30.08.2013
3	Havsa-Osmanlı	41°35'00N 26°50'13E	82m	22.08.2013 11.06.2014
4	Lalapaşa-Sinanköy	41°48'55N 26°41'49E	127m	05.07.2013
5	Lalapaşa-Vaysal	41° 56′ 26N 26° 52′ 9E	438m	05.07.2013
6	Süloğlu -around the Süloğlu Dam	41° 46′ 8N 26° 54′ 36E	155m	21.07.2013
7	Edirne-Center	41°40'33N 26°33'31E	41m	12.06.2014 20.08.2014
8	Enez-Çavuşköy	40°41'17N 26°10'15E	68m	04.08.2008
9	Uzunköprü- Center	41°15'58N 26°41'19E	32m	08.07.2009 15.07.2009
10	İpsala-Center	40°55'16N 26°22'58E	19m	15.07.2009

Table 1. Continued

Table 1.	Continued			
11	Lalapaşa-Çömlekköy	41°50'34N 26°36'38E	101m	14.08.2013
12	Enez- Center	40°43'29N 26°4'57E	24m	04.09.2013
13	Center-Bosnaköy	41°37'35N 26°36'12E	41m	29.05.2014
14	Center-Üyüklütatar	41°32'19N 26°36'3E	44m	29.05.2014
15	Havsa-Bostanlı	41°36'50N 26°58'0E	90m	11.06.2014
16	Havsa-Yolageldi	41°31'0N 26°56'60E	59m	11.06.2014
17	Havsa-Tahal	41° 5'0N 26°51' 0E	42m	13.06.2014
18	Uzunköprü- Değirmenci	41°18'37N 26°42'2E	48m	13.06.2014
19	Uzunköprü-Meşeli	41°22'60N 26°43'60E	83m	13.06.2014
20	Uzunköprü-Başağıl	41°15'49N 26°47'49E	39m	03.07.2014
21	Uzunköprü-Bayramlı	41°18'24N 26°49'34E	77m	03.07.2014
22	Uzunköprü-Çöpköy	41°13'8N 26°49'18E	87m	03.07.2014
23	Uzunköprü-Gazimehmet	41°12'0N 26°55'0E	112m	03.07.2014
24	Uzunköprü-Kırcasalih	41°23'33N 26°48'11E	98m	03.07.2014
25	Uzunköprü-Muhacirkadı	41°,20′,30N 26°,52′,18E	27m	03.07.2014
26	Uzunköprü-Sipahi	41°13'60N 26° 52' 60E	89m	03.07.2014
27	Uzunköprü-Yağmurca	41°10'0N 26°51'0E	125m	03.07.2014
28	Uzunköprü-Çobanpınar	41°10'60N 26° 33' 0E	25m	07.07.2014
29	Uzunköprü-Karapınar	41°7' ON 26°37'60E	50m	07.07.2014
30	Uzunköprü-Kavacık	41°11'14N 26°39'56E	53m	07.07.2014
31	Uzunköprü-Kavakayazma	41°7'60N 26°31'60E	27m	07.07.2014
32	Uzunköprü-Kırköy	41°6'0N 26°43'60E	134m	07.07.2014
33	Uzunköprü-Süleymaniye	41°7'22N 26°48'30E	261m	07.07.2014
34	Lalapaşa-Çömlekakpınar	41°51'0N 26°37'0E	148m	09.07.2014
35	Lalapaşa-Çalıdere	41°55'60N 26°43'60E	197m	15.07.2014
36	Lalapaşa-Kalkansöğüt	41°58'60N 26°48'60E	414m	15.07.2014
37	Lalapaşa-Ömeroba	41°55'0N 26°55'60E	328m	15.07.2014
38	Lalapaşa- Süleymandanişment	41°53'60N 26°53'60E	383m	15.07.2014
39	Süloğlu-Taşlısekban	41°47'60N 26°52'60E	195m	15.07.2014
40	Enez-Arnavutköy	41°43'0N 26°34'60E	76m	04.08.2014
41	Lalapaşa-Kavaklı	41°47'60N 26°45'0E	174m	04.08.2014
42	Meriç-Hasırcı	41°16'0N 26°27'60E	54m	04.08.2014
				24.08.2014
43	Meriç-Karayusuflu	41°15'28N 26°25'55E	24m	04.08.2014

Table 1. Continued

44	Uzunköprü-Çiftlikköy	41°14′0N 26° 37′ 0E	10m	04.08.2014
45	Uzunköprü-Karayayla	41°15'0N 26°34'60E	25m	04.08.2014
46	İpsala-İbriktepe	41°0′35N 26°30′11E	127m	12.08.2014
47	İpsala-Sarıcaali	40°59'60N 26°22'0E	53m	12.08.2014
48	Meriç-Karaağaç	41°11'28N 26°25'25E	23m	12.08.2014
49	Meriç-Küplü	41°6'25N 26°21'7E	12m	12.08.2014
50	Meriç-Subaşı	41°08'60N 26°22'0E	19m	12.08.2014
51	İpsala-Pazardere	40°58'40N 26°34'52E	125m	13.08.2014
52	İpsala-Turpçular	40°56'28N 26°26'15E	27m	13.08.2014
53	Keşan-Beğendik	40°55'57N 26°34'26E	114m	13.08.2014
54	Keşan-Altıntaş	41°00'0N 26°42'60E	185m	23.08.2014
55	Keşan-Kozköy	41°1'0N 26°36'0E	63m	23.08.2014
56	Enez-Abdurrahim	40°38'31N 26°15'25E	40m	24.08.2014
57	Enez-Hasköy	41°38'28N 26°51'31E	102m	24.08.2014
58	Enez-Işıklı	40°43'0N 26°18'0E	13m	24.08.2014
59	Enez-Koyuntepe	40°46'0N 26°20'60E	23m	24.08.2014
60	Enez-Küçükevren	40°38'60N 26°10'0E	41m	24.08.2014
61	Enez-Sultaniçe	40°37'35N 26°9'12E	11m	24.08.2014
62	Enez-Yenikarpuzlu	40°46'0N 26° 12' 0E	17m	24.08.2014
63	Keşan-Çelebi	40°40'41N 26°20'49E	135m	24.08.2014
64	Keşan-Orhaniye	41°43'0N 26°25'60E	29m	24.08.2014
65	Keşan-Bahçeköy	40°46'10N 26°40'32E	45m	26.08.2014
66	Keşan-Boztepe	40°50'60N 26°33' 0E	53m	26.08.2014
67	Keşan-Çeltik	40°40'60N 26°33'0E	117m	26.08.2014
68	Keşan-Karahisar	40°45'0N 26°30'0E	21m	26.08.2014
69	Keşan-Koruklu	40°39'60N 26°25'0E	51m	26.08.2014
70	Keşan-Mecidiye	40°38'20N 26°32'14E	29m	26.08.2014
71	Keşan-Mercan	40°44'60N 26°36'0E	72m	26.08.2014
72	Keşan-Suluca	40°41'20N 26°27'49E	145m	26.08.2014
73	Lalapaşa-Hacıdanişment	41°54'35N 26°49'26E	508m	07.06.2016
74	Kırklareli–Lüleburgaz– Evrensekiz	41°22'11N 27°29'42E	73 m	14.05.2016
75	Kırklareli- between Lüle- burgaz–Pınarhisar	41°29'57N 27°24'51E	106m	16.06.2016

Family: NABIDAE A. Costa, 1853

Subfamily: NABINAE A. Costa, 1853

Genus: NABIS Latreille, 1802

Nabis (Nabicula) flavomarginatus Scholtz, 1847

Material: EDİRNE: İpsala (Turpçular), 13.08.2014, 13; Keşan (Bahçeköy),

26.08.2014, 233; **Uzunköprü** (Karapınar), 07.07.2014, 233

Distribution in Turkey: Turkey Asian part (Anatolia) (Kerzhner, 1996).

Nabis (Nabis) pseudoferus Remane, 1949

Material: EDİRNE: Center-Karaağaç, 12.08.2014, 2♀♀, 7♂♂; Enez- Center 5m 04.09.2013 13; Cavuşköy, 04.08.2008, 1\, 233; Arnavutköy, 04.08.2014, 8\, 533; 24.08.2014, 499, 333; Işıklı, 24.08.2014, 299, 433; 24.08.2014, 10^{\text{Q}}, 1_{\delta}; Sultanice, 24.08.2014, 2^{\text{Q}}, 2_{\delta}; Yenikarpuzlu, 24.08.2014 5\text{\pi}, 4\text{\pi}; **Havsa-**Arpac, 30.08.2013, 8\text{\pi}, 13\text{\pi}; Osmanlı, 22.08.2013, 7\text{\pi}, 1\text{\pi}; Necatiye, 30.08.2013, 533; Bostanlı, 11.06.2014, 1899, 533; Yolageldi, 11.06.2014, 299, 13; Tahal, 13.06.2014, 1799, 333; **İpsala**-Center 15.07.2009, 13; İbriktepe, 12.08.2014, 3\$\partial\$, 2\$\frac{1}{12}\$; Sarıcaali, 12.08.2014, 4\$\partial\$, 5\$\frac{1}{12}\$; Pazardere, 13.08.2014 2\$\partial\$, 9♂♂; Turpçular, 13.08.2014 10♀♀, 8♂♂; Keşan-Beğendik, 13.08.2014, 4♂♂; Kozköy 23.08.2014, 299, 13; Altıntaş, 26.08.2014, 499; Bahçeköy, 26.08.2014, 699, 23; Boztepe, 26.08.2014 13; Çeltik, 26.08.2014, 499; Karahisar, 26.08.2014, 299, 333; Koruklu, 26.08.2014, 1 $\stackrel{\frown}{}$, 1 $\stackrel{\frown}{}$; Mecidiye, 26.08.2014, 14 $\stackrel{\frown}{}$, 7 $\stackrel{\frown}{}$; Mercan, 26.08.2014 299, 233; Orhaniye, 24.08.2014, 599, 533; Suluca, 26.08.2014, 499, 233; **Lalapaşa** -Sinanköy, 05.07.2013, 833, 1599; 05.07.2013, 299,433; Vaysal, 05.07.2013, 13; 14.08.2013, 13; Çömlekakpınar, 09.07.2014, 733; 15.07.2014, 2♀♀, 4♂♂; Kalkansöğüt, 15.07.2014, 3♀♀, 2♂♂; Ömeroba, 15.07.2014 Süleymandanişment, 15.07.2014, $1_{\vec{a}}$; Kavaklı, 04.08.2014, 899, $1_{\vec{a}}$; **Meriç**-Karayusuflu, 04.08.2014, 433; Subaşı, 12.08.2014, 299, 333; Hasırcı, 24.08.2014, 10°, 633; **Süloğlu**- around the Süloğlu Dam. 21.07.2013, 2°, 1133; 21.07.2013 299; Taşlısekban, 15.07.2014, 13♀♀, 7♂♂; **Uzunköprü**-Center, 15.07.2009, 13; Meşeli, 13.06.2014, 19; Başağıl, 03.07.2014, 699, 333; Bayramlı, 03.07.2014, 253, Çöpköy, 03.07.2014, 299, 433; Gazimehmet, 03.07.2014, 1499, 1033; Kırcasalih, 03.07.2014, 399,13; Muhacirkadı, 03.07.2014, 1099, 933; Sipahi, 03.07.2014, 899, 533; Yağmurca, 03.07.2014, 233; Çobanpınar, 07.07.2014, 599, 233; Kavacık, 07.07.2014, 19, 333; Karapınar, 07.07.2014, 4099, 2933; Kavakayazma, 07.07.2014, 1\varphi; Kırköy, 07.07.2014, 7\varphi\varphi, 4\ddots\varphi; S\varphileymaniye, 07.07.2014, 12\ddots\varphi; Karayayla, 04.08.2014, 10♀♀, 4♂♂.

Nabis (Nabis) punctatus A. Costa, 1847

Material: EDİRNE: Center, 12.06.2014, 5, 1; Karaağaç, 12.08.2014, 1; İpsala-Turpçular, 13.08.2014, 1; Meriç-Küplü, 12.08.2014, 4, 4, Uzunköprü-Gazimehmet, 03.07.2014, 1.

Nabis (Nabis) ferus (Linnaeus, 1758)

Material: EDİRNE: **Center**, 12.06.2014, 3\$\$\pi\$, 2\$\$\tau\$; Bosnaköy, 29.05.2014, 6\$\$\tau\$; Üyüklütatar, 29.05.2014, 2\$\pi\$, 5\$\$\tau\$; Karaağaç, 12.08.2014, 1\$, 8\$\$\tau\$; **Enez**-Arnavutköy, 04.08.2014, 1\$\pi\$\$\pi\$\$\tau\$\$\tau\$\$, Abdurrahim, 24.08.2014, 1\$\$\tau\$\$; Çelebi, 24.08.2014, 3\$\pi\$\$\$\tau\$\$\tau\$\$, 2\$\$\tau\$\$\tau\$\$; Hasköy, 24.08.2014, 1\$\$; Işıklı, 24.08.2014, 3\$\pi\$\$\$\$\tau\$\$\tau\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\tau\$\$\tau\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\$\tau\$\$\tau\$\$\$\tau\$\$\

ceköy, 26.08.2014, 299, 13; Boztepe, 26.08.2014, 13; Celtik, 26.08.2014, 299, 233; Karahisar, 26.08.2014, 1 \updownarrow , 1 \circlearrowleft ; Mecidiye, 26.08.2014, 10 \updownarrow \updownarrow , 5 \circlearrowleft \circlearrowleft ; Mercan, 26.08.2014, 1ç, 1ʒ; Suluca, 26.08.2014, 2çç, 2ʒʒ; **Meriç**-Hasırcı, 04.08.2014, 12çç, 1233; **Süloğlu**-Taşlısekban, 15.07.2014, 899, 433; **Havsa**-Bostanlı, 11.06.2014, 10\$\text{\pi}, 6\$\frac{1}{2}\$; Osmanlı, 11.06.2014, 2\$\frac{1}{2}\$\text{\pi}, 15\$\frac{1}{2}\$\text{\pi}\$; Tahal, 13.06.2014, 10\$\frac{1}{2}\$\text{\pi}, 11\$\frac{1}{2}\$\text{\pi}; **Uzunköprü**-Değirmenci, 13.06.2014 5♂; Çöpköy, 03.07.2014, 1♀, 4♂♂; Gazimehmet, 03.07.2014, 10♀♀, 9♂♂; Muhacirkadı, 03.07.2014, 7♀♀, 1♂; Sipahi, 03.07.2014, 6♀♀, 733; Çobanpınar, 07.07.2014, 599, 233; Karapınar, 07.07.2014, 1599, 1633; Çiftlikköy, 04.08.2014, 3♂; Karayayla, 04.08.2014, 4♀♀, 7♂♂.

Distribution in Turkey: Ağrı, Amasya, Antalya, Bursa, Diyarbakır, Elazığ, Erzurum, Giresun, Iğdır, Hatay, Kahramanmaraş, Kars, Tokat (Hoberlandt, 1856; Dursun, 2011; Yıldırım et al., 2013).

Genus: HIMACERUS Wolff, 1811

Himacerus (Aptus) mirmicoides (O. Costa, 1834)

Material: EDİRNE: Center-Sarayiçi (Tavukormanı), 20.08.2014, 1♀.

Subfamily: PROSTEMMATINAE Reuter, 1890

Genus: PROSTEMMA Laporte, 1832

Prostemma sanguineum (Rossi, 1790)

Material: EDİRNE: Lalapaşa-Hacıdanişment, 07.06.2016, 13, 3 nimf; KIRKLAELİ: **Lüleburgaz**-Evrensekiz, 14.05.2016, 1♀, 1♂; between Lüleburgaz-Pınarhisar, 16.06.2016, 233.

RESULTS AND DISCUSSION

In this study, 6 species belonging to 3 genera from Nabinae and Prostemmatinae subfamilies of Nabidae family determined in Turkish Thrace. Among Anatolia is based on quite old years, it is species, Nabis flavomarginatus not very common in Anatolia. Scholtz, 1847 and Nabis ferus (Linnaeus, 1758) are new records for the Heteroptera Fauna of the Thrace Region and additional records of 4 known species from the region were given.

species in Turkey (Kerzhner, 1996; Aukema, the male is pointed in the form of spines. 2020), it is given only as the "Asian part" specifying the locality. species, which has a wide distribution in and Asia in the Palearctic distribution, also spreads in Alaska, Canada, Greenland in the Nearctic Region, apart from the Palearctic.

Nabis (Nabis) ferus has a wide distribution, and was detected in 37 different localities in Edirne Province in this study.

were Although its known distribution from

In the Palearctic Region, it shows a very wide distribution in Europe and Asia, including Turkey and the neighboring countries to the research area (Kerzhner, 1996). This species was recorded for the The first findings of Nabis flavomarginatus first time in the Thrace Region during this for the Thrace Region, was detected in 3 study. It is very similar to N. pseudoferus localities located in the south of Edirne and N. punctatus species in terms of (Uzunköprü - Karapınar, İpsala - Turpçu- general appearance, but it is easily lar, Kesan - Bahçeköy). When taking into distinguished from these two species by account the distributional data of the the fact that the apex of the paramer in

> The other 4 species, Nabis pseudoferus, N. punctatus, Himacerus mirmicoides and Prostemma sanguineum identified in this study, arevery common both in Turkey and intheir general geographical distribution. N. pseudoferus was also detected in many localities during the research.

The species of the genus Nabis were generally detected in the alfalfa fields or the wild green grasses on the field or Dursun A., Fent, M., 2015, Notes on some roadside during our research. habitats, where the species are detected, are generally rich in other soft-bodied insect larvae or adults hunted by the species. It is thought that Nabis species chose these habitats because of the Erbay, H., 1986, Türkiye Nabidae (Heteroptera) convenience in finding prey by them. Prostemma sanguineum was found on the ground in the alfalfa fields, and under stones and Himacerus mirmicoides in the mixed forest dominated by poplar.

In the study, when the most common species in 75 localities where field studies Gadeau De Kerville, H., 1939, Voyage zoologique were carried out were evaluated respectively; Nabis pseudoferus was detected from 62 localities, N. ferus from 37 localities, N. punctatus from 5 localities, N. flavomarginatus Hoberlandt, L., 1956, Results of the Zoological and Prostemma sanguineum from 3 localities, and Himacerus mirmicoides from only 1 locality.

When the data obtained as a result of the research were evaluated, the number of species of Nabidae family in the Thrace Region increased to a total of 12 with the addition of two new records. In addition, new data on the distribution of previously known species were obtained.

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First record of *Dryadocoris apicalis* (Herrich-Schaeffer, 1842) (Hemiptera: Heteroptera: Pentatomidae) in Greece

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ABSTRACT: The first record of Dryadocoris apicalis (Herrich-Schaeffer, 1842) for Greece is reported. Information on the known distribution of the species is summarized.

KEYWORDS: Dryadocoris apicalis, first record, distribution, Greece.

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France (including Corsica), (including Capraia, Pantelleria, Sardinia 2007; Robertson, 2009). and Sicily), Portugal and Spain (including the Canary Islands) (Carapezza, 1995; Jiménez et al., 2003; Aukema et al., 2006; Dusoulier & Lupoli, 2006; Rider, 2006; Belousova, 2007; Gessé, 2011; Roca-Cusachs & Goula, 2017; Fernández Ruiz, 2018; Roca-Cusachs et al., 2018; The specimen was found inside a building van der Heyden, 2019).

Outside Europe, the species has been reported from Algeria, Cameroon, the

So far, Dryadocoris apicalis (Herrich- Democratic Republic of the Congo, the Schaeffer, 1842) (Hemiptera: Heteroptera: Republic of the Congo, Eritrea, Ethiopia, Pentatomidae) has been reported from Guinea, Kenya, Morocco, Nigeria, Rwanda, the following European countries: Albania, Somalia, South Africa, Sudan, Tanzania, Italy Togo and Yemen (Rider, 2006; Belousova,

> Now, the first record of D. apicalis in Greece can be reported: On 02.11.2021, an adult specimen was photographed by Christopher Bakos in Parga, a coastal village in North-West Greece (Fig. 1).

> where vegetable plants are grown hydroponically. Likely, it was drawn to the heat of the LED lamp that provides the sunlight/heat to the vegetables

alternatively to the moisture at the base ACKNOWLEDGEMENTS of the vegetable plant (Lettuce) where the bug was found (Christopher Bakos, pers. comm.). So far, the nearest location of findings of other specimens of *D. apicalis* is the Albanian city of Vlorë, located less than 200 km north-west of Parga (van der Heyden, 2019).

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Figure 1. Dryadocoris apicalis (Herrich-Schaeffer, 1842), Parga, Greece, 02.11.2021. (Photo: Christopher Bakos).



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Some Heteroptera (Hemiptera) species that are potential natural enemies of *Cimbex quadrimaculata* (Müller, 1766) (Hymenoptera: Cimbicidae)

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ABSTRACT: The study was carried out in the provinces of Elazig and Diyarbakir between 2020-2021. In the studies conducted in nature and in laboratory conditions, it is observed that the species does not feed on advanced larval stages of the pest. From the species identified in the study, *Deraeocoris trifasciatus* Linnaeus, 1767 and *Globiceps sphaegiformis* Rossi, 1790 species belonging to the Miridae family were observed in nature, and *Nagusta goedelii* Kolenati, 1857 and *Zelus (Diplodacus) renardii* (Kolenati, 1856) species belonging to the Reduviidae family were observed to in laboratory conditions to feed on the pest. *Geocoris luridus* Fieber, 1844 and *Geocoris putonianus* Bergroth, 1892 species belonging to the Geocoridae family were found to be abundant in nature before the pest began to be observed in nature. *Brachycoleus steini* Jakovlev, 1884 specimens and *Macrotylus ponticus* Seidenstücker, 1966 found in abundance in the garden at the larval period of pest of these species.

It has not been determined that they feed on pests. All species have been detected in the fauna of both provinces. These species are also the first record for the provinces where they have been found.

KEYWORDS: Almond, *Cimbex quadrimaculata*, Natural Enemies, Predator.

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INTRODUCTION

The Cimbicidae is a small family, with 196 species occurring in the World (Özbek, 2014). In Turkey; There were a 23 species (Önder, 2011). Cimbex quadrimaculata

(Müller, 1766) is an important pest in fruit-growing areas that especialy on almonds of Turkey (Bolu, 2016) (Figure 1). *C. quadrimaculata* was very serious pest on young almond plants (Özbek,



2014; Çakıcı et al., 2015). When this pest Opheltes glaucopterus (Linnaeus) and significant damage of many fruits.

Listrognathus mactator for the first time. Özbek (2014) obtained order of the pest.

is not methodological managed, it creates *Phobetes nigriceps* (Gravenhorst) (Hymenoptera: Ichneumonidae: Ctenopelmatinae), as larva Additionaly; There is no licensed chemical – pupa parasitoids of C. quadrimaculata. control recommended against to this P. nigriceps was obtained for the first time species. Therefore, its biological control is from C. quadrimaculata larvae. No any an opportunity. Özgen et al. (2010) obtained studies have been conducted on the pre-(Thunberg) dator species of this species. The aim of (Hymenoptera: Ichneumonidae: Cryptinae) the study is to determine the predator as a parasitoid of C. quadrimaculata species belonging to the Heteroptera





Figure 1. The habitus of Cimbex quadrimaculata larvae and feeding on almond trees.

MATERIAL AND METHODS

Elazığ and Diyarbakır provinces in culture plates. Species that have been Anatolia Region of Turkey in 2021-2022. found to feed in nature and the biological The studies were carried out between periods of the pest have been recorded. 03.05.2021 and 15.06.2021 in Diyarba- Miridae species could not be brought kır, Egil and Elazığ province and Keban alive from natural conditions to laboratory district. Studies were carried out in a conditions, while Reduviidae species climate cabinet with 65% humidity and brought from nature were left in the (Figure 2). When the larvae started to plates. It has been observed that some appear in nature, 50 1st instar larvae species insert their stylets into the pest.

were collected and brought to the laboratory The present study was carried out in and left to feed on almond leaves in ± 2°C temperature conditions same environment as pests with culture





Figure 2 a) Climate Cabinet b) Containers with Cimbex quadrimaculata larvae and pupa.

RESULTS and DISCUSSION

Following 8 species were found in the C. quadrimaculata research conducted in two localities in Elazığ and Diyarbakır (Table 1)

Table 1. Heteroptera species, Localities and Biological Observations

Family	Species	Locality	Biological Stage of Cimbex quadrimaculata larvae fed by the predator (*Field and labortory, **Field,***Potential)
MIRIDAE	Brachycoleus steini Jakovlev, 1884	Diyarbakır, Eğil, Elazığ, Keban	No feeding (Abundant)
MIRIDAE	Deraeocoris trifasciatus Linnaeus, 1767 **	Diyarbakır, Eğil, Elazığ, Keban	1st instar larvae
MIRIDAE	Globiceps sphaegiformis Rossi, 1790 **	Diyarbakır, Eğil, Elazığ, Keban	1 instar larvae
MIRIDAE	<i>Macrotylus ponticus</i> Seidenstücker, 1966	Diyarbakır, Eğil, Elazığ, Keban	No feeding (Abundant)
REDUVIIDAE	<i>Nagusta goedelii</i> Kolenati, 1857 *	Diyarbakır, Eğil, Elazığ, Keban	1st instar larvae.
REDUVIIDAE	Zelus (Diplodacus) renardii (Kolenati, 1856) *	Diyarbakır, Eğil	1st instar larvae.
GEOCORIDAE	Geocoris luridus (Fieber, 1844)***	Diyarbakır, Eğil, Elazığ, Keban	Before the pest occured in nature, it was observed in each orchards in many ammounts.
GEOCORIDAE	Geocoris putonianus Bergroth, 1892 ***	Diyarbakır, Eğil, Elazığ, Keban	Before the pest occured in nature, it was observed in each orchards in many ammounts.

The species has been seen during the months of March and July. They are abundant in the egg and larval stage of the pest (Figure 3).

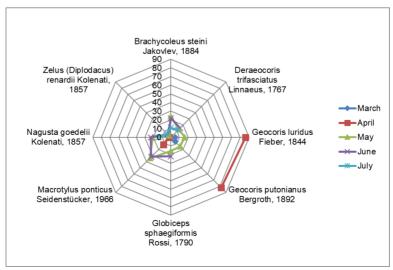


Figure 3. Dates of Predator Species in Nature

were observed in field conditions where Nagusta goedelii Kolenati, 1857 species they were fed in the first larval stage of do not feed on the biological periods of the pest. Since the species belonging to the pest after the 2nd larval period. For this family could not be brought to the Brachycoleus steini; The presence of the laboratory, conditions alive, the culture Centaurea sp. plant in the garden edges plate feeding experiments could not be has also increased the probability of this conducted. In these species; Globiceps species being found in almond trees. sphaeqiformis Rossi, 1790 was determined Abundant varieties of this plant in this in the provinces of Elazığ and Divarbakır species in the literature are supportive in previous years (Matocq et al., 2014; (Hosseini, 2016). Phytophagous status in Özgen and Dioli, 2019). There are no Macrotylus species should be investigated. studies on the food preferences of the species belonging to this genus. Kıyak ACKNOWLEDGEMENTS (2020); reported that Deraeocoris spp. is zoophytophageous, generally feeding on This study was supported by TUBITAK aphids and it preys on eggs of true bugs, 1180124 number" 3001 project" and All larvae of whiteflies. However, there is no record of it feeding on the Hymenopera On the other hand; species. (Diplodacus) renardii (Kolenati, 1856) and Nagusta goedelii (Kolenati, 1857) species collected from the field and brought to Bolu, H., 2016, Distrtibution, Life History and the laboratory were fed with the first larval stage of the pest. These species stand out as the polyphagous predator. Celik et al., 2021; reported that this species is fed on Allantus (s.str.) viennensis (Schrank, 1781) in Diyarbakir. N. goedelii generalist predator potentially capable of attacking the eggs and first instar nymphs of some Pentatomidae specimens and gypsy moth (Lymantria dispar (Linnaeus, 1758), Lepidoptera (Erebidae) (Bigsby et al., 2011; Bulgarini et al., 2020). The feeding record on this species is the first. Geocoris spp. are among the most important predaceous insects in the world. They feed on eggs and small larvae of most lepidopteran pests. Moreover, Geocoris sp. species were found to be abundant in almond trees in gardens where they formed a population before the pest began to appear in nature. In studies conducted in Çelik, H., Dioli, P., Bolu, H., 2021, First nature and laboratory conditions, it was found that the species did not feed on the advanced larval periods of the pest. All species have been detected in the fauna of both provinces. These species are also the first record for the provinces where they have been found. It has been observed

Species belonging to the Miridae family that Zelus (Diplodacus) renardii

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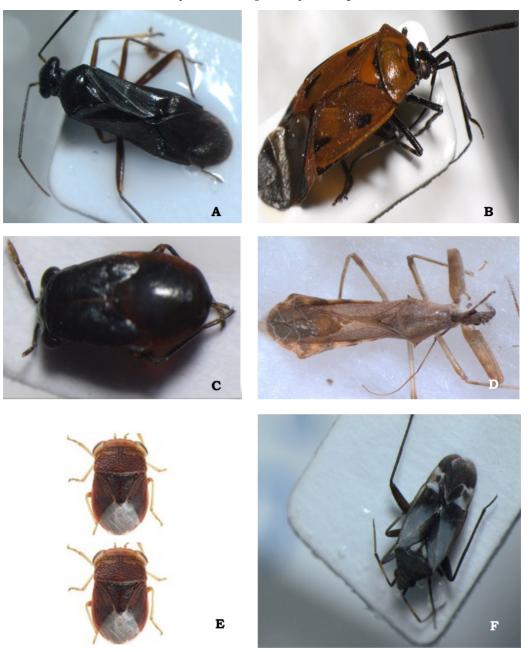
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Figues 2. The Figures of Heteroptera Species in Almond Orchards. A: Globiceps sphaegiformis Rossi, 1790; B: *Deraeocoris trifasciatus* Linnaeus, 1767; C: *Geocoris* sp. nymph, D: *Nagusta goedelii* Kolenati, 1857; E: *Geocoris putonianus* Bergroth, 1892 (Çerçi and Özgen, 2021); F: *Macrotylus ponticus* Seidenstücker, 1966.



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A study on the Coreoidea (Hemiptera: Heteroptera) fauna of Çorum Province

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ABSTRACT: In this study is reported28 species belonging to 20 genera from 4 families (Alydidae, Coreidae, Rhopalidae and Stenocephalidae) of the superfamily Coreoidea on the material collected from 39 different localities in Çorum province between 2019 and 2021. Of them, 23 species are new records for the Coreoidea fauna of Çorum. While 20 species recorded in the present study are of Holomediterranean origin, 3 species of Pontomediterranean origin, 2 species of Irano-Turanian origin, 1 species has distribution in Euro-Siberian, Mediterranean and Nearctic. As a result of our observations in the field, a total of 21 taxon at genus level from 15 plant families were identified in relation to Coreoidea species.

KEYWORDS: Heteroptera, Alydidae, Coreidae, Rhopalidae, Stenocephalidae, new records, Çorum, Turkey.

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INTRODUCTION

The superfamily Coreoidea take part in the infraorder Pentatomomorpha (Hemiptera: Heteroptera) and that includes families

Alydidae, Coreidae, Rhopalidae and Stenocephalidae in Palaearctic region. Currently approximately 344 species in 84 genera of family Coreidae, approximately



69 species in 26 genera of family climate is generally dominant in the Alydidae, 71 species belonging to 14 province. The Black Sea climate type is genera of family Rhopalidae and 18 spe- especially prevails in Bayat, Dodurga, cies of a genus of family Stenocephalidae are İskilip, Kargı, Laçin, Oğuzlar, Osmancık Palearctic the (Dolling, 2006). Data studies on Coreoidea in Turkey, show Mecitözü, Ortaköy and Sungurlu districts that there are 7 species in 4 genera of prevails Central Anatolian climate. The Alydidae, 48 species from 20 genera of winter months are cloudy and the Coreidae, 29 species in 11 genera of summers are hot with the effect of the Rhopalidae, and 7 species in 1 genus of Central Anatolian climate. Stenocephalidae. Of those, Camptopus southern parts of İskilip is the steppes bifasciatus Fieber, 1864 of the family consisting of herbaceous plants. Corum Alvdidae from Amasya and Anoplocerus province is located at the intersection of luteus (Fieber, 1861) from İzmir, Arenocoris Irano-Turanian and Euro-Siberian plant latissimus Seidenstücker, Ulukısla (Niğde), Cercinthus (Fieber. region, Ceriomeris armeniacus Tshernova, flora and faunal elements (Kaya, 2015). 1978 from easten Anatolia of the family The aim of this study is to give the Coreidae, Corizus brevicornis Horváth, current fauna of superfamily Coreoidea, 1917 from Oltu (Erzurum), Corizus fenestella and to open a new way for scientific and Horváth, 1917 from Tuzluca (Iğdır) of the ecological studies in Corum province. family Rhopalidae and Dicranocephalus 1897) from Hassa MATERIAL AND METHODS putoni (Horváth, (Hatay) of the family Stenocephalidae are

The study material was obtained from 39 Noualhier, 1895; Horváth, 1883, 1901; localities with different vegetation and Fahringer, 1922; Kiritshenko, 1924; Hoberlandt, 1955; Seidenstücker, from 2019 to 2021 (Fig. 1). 1958, 1960; Wagner, 1959, 1966); Linnavuori, The specimens were collected from 1965; Pehlivan, 1981; Kıyak, 1990a, b, c; herbaceous vegetation with a sweep net 1993; 2000; Moulet, 1995; Özsaraç & Kıyak, 2001; Önder et al., 2006; Dursun, 2009, 2011; Dursun & Fent, 2009, 2017; Dursun et al., 2010; Fent & Kment, 2011; Fent & Dursun, 2019 and Zengin laboratory. In the laboratory, specimens & Dursun, 2019).

feeding on fecal and putrefaction matters and that humans were bitten by species of superfamily Coreoidea, especially the species are phytophagous and feed on meristematic tissues and ripe seeds. Many species of Coreoidea are known as economically and biologically important (Dolling, 2006; Faúndez, 2016).

The study area Corum is located in the Central Black Sea region and is a transition area between the Black Sea and the Central Anatolia region. The Black Sea

region and Uğurludağ districts situated in the based on the Kızılırmak basin. In Alaca, Boğazkale, 1960 from belt. Therefore Corum province has very griseus rich areas in terms of microclimate. 1861) from Turkish Thrace different vegetation and habitat properties of

1918, habitat in Corum province in the years

and from trees plants with a Japanese umbrella. All samples were put in tubes in 70% ethanol and brought to the were softened in hot water (80°C-100°C) Although it has been rarely reported of for preparation of the male genitalia which was used for further identifications. The specimens were prepared and identified relevant diagnostic using the investigated under a stereomicroscope (Leica EZ4) and keys of Stichel (1960), Pehlivan (1981) and Moulet (1995).

The material is deposited in the collection of Amasya University, Faculty of Science and Arts, Department of Biology (Amasya, Turkey).

RESULTS

Hemiptera Linnaeus, 1758 Heteroptera Latreille, 1810 Coreoidea Leach, 1815 Alydidae Amyot & Serville, 1843 Alydinae Amyot & Serville, 1843

Alydus Fabricius, 1803

Alydus calcaratus (Linnaeus, 1758)

Material examined: Çorum: Center, 19.07.2020, 299, 233; Çatak, 03.07.2021, 133;

Osmancık: Yaylabaşı, 06.10.2020, 13; Sarpınkavak, 19.10.2020, 200.

Comments: This Holomediterranean species was found under *Astragalus* sp. and it is widespread and frequently distributed in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Fent & Dursun, 2019; Zengin & Dursun, 2019). In the study, this species was recorded for the first time from Çorum province.

Camptopus Amyot & Serville, 1843

Camptopus lateralis (Germar, 1817)

Material examined: Osmancık, Center, 25.05.2019, 1_{\circ} ; 23.04.2021, 1_{\circ} ; Sarpınkavak, 19.10.2020, 1_{\circ} , 1_{\circ} ; **Alaca**, 05.06.2021, 1_{\circ} ; **Kargı**, Karapürçek, 02.10.2021, 1_{\circ} , 9_{\circ} .

Comments: This Holomediterranean species was found on *Chrysantemum* sp., *Rosa* sp., and *Rumex* sp. and it is widespread and frequently distributed in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Fent & Dursun, 2019; Zengin & Dursun, 2019; Çerçi et al., 2016; Çerçi & Özgen, 2021). In the study, this species was recorded for the first time from Çorum province.

Camptopus tragacanthae (Kolenati, 1845)

Material examined: Osmancık, Akören Plateau, 18.04.2020, 1_{\circ} ; Sarpınkavak, 19.10.2020, 1_{\circ} .

Comments: This Irano-Turanian species was found under *Astragalus* sp. and it is widespread and frequently distributed in Turkey (Pehlivan, 1981; Kıyak et al., 2004; Önder et al., 2006; Dursun, 2009; Kıyak & Akar, 2010; Fent & Japoshvili, 2013; Küçükbasmacı & Kıyak, 2015; Kıyak, 2016; Özgen et al., 2018; Fent & Dursun, 2019; Dursun, 2009; Kıyak & Baş, 2021). In the study, this species was recorded for the first time from Çorum province.

Coreidae Leach, 1815 Pseudophloeinae Stål, 1868

Ceraleptus A. Costa, 1847

Ceraleptus gracilicornis (Herrich-Schaeffer, 1835)

Material examined: Kargi, Uzunyurt, 19.06.2021, 13.

Comments: This Holomediterranean species is rarely encountered in Turkey and it was recorded for the first time from Çorum province(Önder et al., 2006; Baş, 2013; Yıldırım et al., 2013; Seferoğlu, 2013; Küçükbasmacı & Kıyak, 2015; Fent & Dursun, 2019; Zengin & Dursun, 2019). It has been found on *Trifolium* sp. in the study.

Coriomeris Westwood, 1842

Coriomeris affinis (Herrich-Schaeffer, 1839)

Material examined: Kargi, Karapürçek, 20.06.2020, 233.

Comments: The known distribution of this Mediterranean species in Turkey is widely and frequently distributed (Puton, 1892; Seidenstücker, 1960; Önder et al., 2006; Baş, 2013; Matocq, 2014; Çerçi et al., 2016; Fent & Dursun, 2019; Zengin & Dursun, 2019). In the study, this species was recorded for the first time from Çorum province and it has been found on *Pinus* sp.

Coriomeris denticulatus (Scopoli, 1763)

Material examined: Alaca, 05.06.2021, 399, 333; Osmancık, Öbektaş Plateau, 24.07.2021, 13.

Comments: The known distribution of this Euro-Siberian species is common and widely distributed in Turkey (Horváth, 1883; Hoberlandt, 1955; Linnavuori, 1965; Önder et al., 2006; Dursun, 2011; Fent & Dursun, 2019; Zengin & Dursun, 2019; Bolu, 2020; Bulak Korkmaz & Yıldırım, 2021). This species was found on *Lamium* sp. and the record of the species given in the present study is for the first time from Çorum province.

Coreinae Leach, 1815

Leptoglossus Guérin-Méneville, 1831

Leptoglossus occidentalis Heidemann, 1910

Material examined: Çorum, Center, 17.08.2020, 1° , 1_{\circ} ; Osmancık, Yaylabaşı, 04.05.2020, 1° , 1_{\circ} ; Sarpınkavak, 19.10.2020, 1_{\circ} ; Laçin, Yeşilgöl, 03.10.2021, 1° .

Comments: This western conifer seed bug is an invasive alien species originated from North America. The distribution of *Leptoglossus occidentalis* rapidly expanded to the eastern and western of Turkey. This species was found on *Pinus* sp. This Neactic species is common and widely distributed in Turkey (Arslangündoğdu & Hızal, 2010; Fent & Kment, 2011; Hızal & İnan, 2012; Çerçi & Koçak, 2016; Özgen et al., 2017; Parlak, 2017; Yücel & Kıvan, 2018; Zengin & Dursun, 2019; Oğuzoğlu & Avcı, 2020).

Centrocoris Kolenati, 1845

Centrocoris spiniger (Fabricius, 1781)

Material examined: Çorum, Center, 06.06.2021, 1; Ortaköy, Mollahasan, 26.07.2020, 1; Osmancık, Sarpınkavak, 19.10.2020, 1.

Comments: This Holomediterranean species is common and widely distributed in Turkey according to available data (Horváth, 1883; Kiritshenko, 1918; Hoberlandt 1955; Wagner, 1966; Tuatay et al., 1972; Kıyak, 1993; Önder et al., 2006; Dursun & Fent, 2009; Dursun, 2011; Fent & Dursun, 2019; Zengin & Dursun, 2019). This species was found on *Cirsium* sp. and it is reported for the first time from Çorum province in the study.

Centrocoris variegatus Kolenati, 1845

Material examined: Osmancık, 23.06.2021, 599, 333.

Comments: This Holomediterranean species is frequently and widely distributed in Turkey according to the available records (Horváth, 1901; Hoberlandt, 1955; Kıyak, 1990a; Kıyak, 1990b; Özsaraç, 2004; Önder et al., 2006; Dursun, 2011; Yıldırım et al., 2011; Fent & Dursun, 2019; Zengin & Dursun, 2019; Bolu, 2020; Koçakoğlu, 2020; Bulak Korkmaz & Yıldırım, 2021). This species was found on *Cirsium* sp. and it is reported for the first time from Çorum province in the study.

Coreus Fabricius, 1794
Coreus marginatus (Linnaeus, 1758)

Material examined: Çorum, Center, 27.06.2020, 1♀, 1♂; 19.07.2020, 2♀♀; 17.08.2020, 1♂; 18.08.2020, 2♀♀; Çatak, 12.06.2021, 5♀♀, 7♂♂; 03.07.2021, 1♀; Kıvırcık, 26.07.2020, 2♀♀, 1♂; Bayat, 19.07.2020, 2♀♀, 2♂♂; Dodurga, Berkköy, 18.09.2020, 1♀, 1♂; İskilip, Seyirtepe, 11.07.2020, 4♀♀; Kargı, Hacıhamza, 02.10.2021, 1♀, 1♂; 26.06.2021, 1♀, 3♂♂; Uzunyurt, 19.06.2021, 3♀♀, 4♂♂; Karapürçek, 02.10.2021, 2♀♀; Mecitözü, Elvançelebi, 30.07.2020, 2♀♀, 2♂♂; Hıdırlık, 09.10.2021, 2♀♀, 3♂♂; 09.10.2021, 4♀♀, 2♂♂; Oğuzlar, Obruk Barajı, 28.09.2020, 1♀; Ortaköy, İncesu; 26.07.2020, 1♀; Oruçpınar, 26.07.2020, 1♂; Mollahasan, 26.07.2020, 1♀, 1♂; Osmancık, 25.05.2019, 1♀; Ardıç, 02.10.2021, 1♀; Akören Plateau, 18.06.2020, 1♂; Yaylabaşı, 04.08.2020, 2♀♀, 3♂♂; 04.05.2020, 1♂; Öbektaş Platau, 24.07.2021, 1♀; Sungurlu, Karakaya, 15.07.2020, 1♀; Koparan, 15.07.2020, 1♀, 1♂; Uğurludağ, 10.07.2021, 1♀.

Comments: This Holomediterranean species is frequently and widely distributed in all the regions of Turkey according to the available records (Hoberlandt, 1955; Önder et al., 2006; Yıldırım et al., 2011; Baş, 2013; Küçükbasmacı & Kıyak, 2015; Fent &Dursun, 2019; Kaçar, 2019; Zengin & Dursun, 2019; Bulak Korkmaz & Yıldırım, 2021; Kıyak & Baş, 2021). This species was found on *Rumex* sp.

Enoplops Amyot & Serville, 1843 Enoplops disciger (Kolenati, 1845)

Material examined: Osmancık, 25.05.2019, 1♀; Ortaköy, Fındıklı, 26.07.2020, 1♂. Comments: This Pontomediterranean species is commonly and widely distributed in Turkey according to available records (Horváth, 1901; Kiritshenko, 1918; 1924; Hoberlandt, 1955; Linnavuori, 1965; Moulet, 1995; Kıyak, 2000; Özsaraç & Kıyak, 2001; Özsaraç, 2004; Önder et al., 2006; Dursun & Fent, 2009; Dursun, 2011; Yıldırım et al., 2011; Zengin & Dursun, 2019). This species was found on *Echium* sp. and it is reported for the first time from Çorum province in the study.

Spathocera Stein, 1860

Spathocera lobata (Herrich-Schaffer, 1840)

Material examined: Corum, Center, 06.06.2021, 1.

Comments: This Holomediterranean species is rarely encountered in Turkey according to available records (Önder et al., 2006; Yıldırım et al., 2013; Bayrakçı, 2011; Dursun, 2011). This species was recorded fort he first time from Çorum province in the sudy and it was found on *Rumex* sp.

Syromastus Berthold, 1827

Syromastus rhombeus (Linnaeus, 1767)

Material examined: Osmancık, 25.05.2019, 1 \updownarrow ; Akören Plateau, 18.04.2020, 1 \updownarrow , 1 \circlearrowleft ; İskilip, Seyirtepe, 11.07.2020, 1 \updownarrow .

Comments: This Holomediterranean species is common in Turkey accordint to evailable records (Horváth, 1883; 1901; Kiritshenko, 1918; 1924); Hoberlandt, 1955; Linnavuori, 1965; Tuatay et al., 1972; Kıyak, 1990a, b; Çağlar, 1992; Özsaraç, 2004; Önder et al., 2006; Dursun & Fent, 2009; Dursun, 2011; Fent & Dursun, 2019; Altınok & Muştu, 2019; Zengin & Dursun, 2019). This species was found on *Poa* sp.

Gonocerus Berthold, 1827

Gonocerus acuteangulatus (Goeze, 1778)

Material examined: **Çorum**, Center, 19.07.2020, $1\capproptial$; Kıvırcık, 26.07.2020, $2\capproptial$; **Laçin**, Yeşilgöl, 01.09.2019, $1\capproptial$; Kırkdilim, 01.09.2021, $1\capproptial$; **Osmancık**, $1\capproptial$; Yaylabaşı, 14.07.2019, $1\cappoonup$; 04.05.2020, $1\capproptial$, 1 \capproptial ; Akören Plateau, 18.04.2020, $1\cappoonup$, 1 \cappoonup . **Comments:** This Holomediterranean species is commonly and widely distributed in

Anatolia according to available records (Horváth, 1883; Hoberlandt, 1955; Linnavuori, 1965; Tezcan & Önder, 1999; Tezcan & Önder, 1999; Özsaraç, 2004; Önder, et al. 2006; Dursun, 2011; Yıldırım et al. 2011; Küçükbasmacı & Kıyak, 2015; Zengin & Dursun, 2019; Bulak Korkmaz & Yıldırım, 2021; Çerçi & Özgen, 2021). This species was found on *Juniperus* sp. and it is reported for the first time from Çorum province in the study.

Gonocerus juniperi Herrich-Schaeffer, 1839

Material examined: Dodurga, Berkköy, 18.09.2020, 1♀, 1♂; Oğuzlar, Obruk Barajı, 28.09.2020, 1♀; Osmancık, Akören Plateau, 18.04.2020, 1♂; Sarpınkavak, 19.10.2020, 1♂; Yaylabaşı, 14.07.2019, 1♀; 06.10.2020, 1♀, 1♂; 08.02.2020, 1♀, 1♂. Comments: This Holomediterranean species is frequent and widely distributed in Anatolia and Turkish Thrace according to the available records (Horváth, 1883; Kiritshenko, 1918; Hoberlandt, 1955; Kıyak, 1993; Önder et al., 2006; Dursun & Fent, 2009; Dursun, 2011; Kıvan & Dirik, 2016; Özgen et al., 2018; Sert & Özdemir, 2019). It was found on *Juniperus* sp. and is reported for the first time from Çorum province in the study.

Phyllomorpha Laporte, 1833

Phyllomorpha laciniata (Villers, 1789)

Material examined: Bayat, 19.07.2020, 1, 1; Dodurga, Berkköy, 18.09.2020, 1, 1; Osmancık, Akören Plateau, 18.04.2020, 1; **Osmancık,** 25.05.2019, 1; **Ortaköy,** Fındıklı, 26.07.2020, 1; Sarpınkavak, 19.10.2020, 1; Yaylabaşı, 14.07.2019, 1.

Comments: *Phyllomorpha laciniata* (=*Phyllomorpha lacerata* Herrich-Schaeffer, 1835) is Holomediterranean species. It is frequent and widely distributed in Anatolia and Turkish Thrace according to the available records (Fahringer, 1922; Hoberlandt, 1955; Kıyak, 1990a; Kıyak, 1990b; Moulet, 1995; Özsaraç, 2004; Önder et al., 2006; Dursun & Fent, 2009; Dursun, 2011; Zengin & Dursun, 2019). This species was found under *Acantholimon* sp. and *Astragalus* sp. and is reported for the first time from Çorum province in the study.

Rhopalidae Amyot & Serville, 1843 Rhopalinae Amyot & Serville, 1843

Brachycarenus Fieber, 1860

Brachycarenus languidus (Horváth, 1891)

Material examined: Çorum, Center, 19.07.2020, 2° , 1_{\circ} ; Kıvırcık, 26.07.2020, 2° , 2_{\circ} ; **Laçin**, Yeşilgöl, 01.09.2019, 1_{\circ} , Kırkdilim, 01.09.2021, 1_{\circ} ; **Osmancık,** 1_{\circ} , 1_{\circ} ; Yaylabaşı, 14.07.2019, 2_{\circ} , 1_{\circ} ; 04.05.2020, 1_{\circ} , 1_{\circ} .

Comments: This Irano-Turanian species is rare in Turkey according to available records (Horváth, 1891; Kiritshenko, 1918; Zengin & Dursun, 2019). This species is reported for the first time from Çorum province in the study and it was found on *Chenopodium* sp.

Brachycarenus tigrinus (Schilling, 1829)

Material examined: Kargı, Karapürçek, 02.10.2021, 200.

Comments: This Holomediterranean species is widespread distributed and frequently found in Turkey (Puton & Noualhier, 1895; Horvath, 1901, 1905; Kiritshenko, 1918; Gadeau de Kerville, 1939; Tuatay et al., 1972; Pehlivan, 1981; Yıldırım et al., 2011; Dursun, 2009; Fent & Dursun, 2019; Zengin & Dursun, 2019; Kaçar, 2019; Bulak Korkmaz & Yıldırım, 2021). This species was found on *Chenopodium* sp.

Corizus Fallén, 1814

Corizus hyoscyami (Linnaeus, 1758)

Material examined: Alaca, 05.06.2021, 1_{\circ} ; Bayat, 19.07.2020, 2° , 1_{\circ} ; Osmancık, 25.05.2019, 1_{\circ} ; 23.04.2021, 1_{\circ} ; 23.06.2021, 2° ; Yaylabaşı, 17.04.2020, 1_{\circ} , 1° ; Akören Plateau, 18.04.2020, 1_{\circ} , 1° ; Sungurlu, 16.10.2021, 1_{\circ} ; Uğurludağ, 10.07.2021, 1_{\circ} .

Comments: This Holomediterranean species is widespread distributed and frequently found in Turkey. (Pehlivan, 1981; Yıldrım et al., 2011; Yücel & Kıvan, 2018; Fent & Dursun, 2019; Zengin & Dursun, 2019; Bolu, 2020; Çerçi & Özgen, 2021; Kıyak & Baş, 2021). This species has been found on the *Hypericum* sp. and *Onosma* sp. It is reported for the first time from Çorum province in this study.

Liorhyssus Stål, 1870

Liorhyssus hyalinus (Fabricius, 1794)

Material examined: Çorum, Center, 23.08.2021, 1, 1; Kargı, Karapürçek, 02.10.2021, 1; Osmancık, Yaylabaşı, 14.07.2019, 1.

Comments: This Holomediterranean and cosmopolitan species is widespread and frequently distributed in Turkey (Pehlivan, 1981; Özsaraç & Kıyak, 2001; Önder et al., 2006; Dursun, 2009; Yıldırım et al., 2011; Matocq et al., 2014; Altınok & Muştu, 2019; Fent & Dursun, 2019; Dursun ve Zengin, 2019; Bulak Korkmaz & Yıldırım, 2021). This species is new record for the Heteroptera fauna of Çorum province and it was found on the *Poa* sp.

Maccevethus Dallas, 1852

Maccevethus corcicus corsicus Signoret, 1862

Material examined: Çorum, Center, 18.10.2020, 1\overline{?}; **Sungurlu,** 30.10.2021, 1\overline{?}.

Comments: The distribution of this Ponto-Mediterranean species is common in the Turkey. (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Fent & Dursun, 2019; Altınok & Muştu, 2019; Zengin & Dursun, 2019; Bulak Korkmaz & Yıldırm, 2021). In the study, this subspecies was recorded for the first time from Çorum. It has been found on the *Poa* sp.

Maccevethus errans caucasicus (Kolenati, 1845)

Material examined: Sungurlu, 16.10.2021, 13.

Comments: This Ponto-Mediterranean subspecies was recorded on the *Erodium* sp. from Çorum for the first time in the study. It is widespread and frequently distributed in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Altınok & Muştu, 2019; Zengin & Dursun, 2019; Bolu, 2020; Çerçi & Özgen, 2021; Kıyak & Baş, 2021).

Rhopalus Schilling, 1827

Rhopalus parumpunctatus Schilling, 1829

Material examined: Çorum, Center, 19.07.2020, $3\stackrel{\frown}{,} 2\stackrel{\frown}{,} 3$; 06.06.2021, $2\stackrel{\frown}{,} 1\stackrel{\frown}{,} 3$; **Kargı,** Karapürçek, 02.10.2021, $2\stackrel{\frown}{,} 9$; **Osmancık,** Ardıç, 02.10.2021, $2\stackrel{\frown}{,} 9$; Yaylabaşı, 17.04.2020, $1\stackrel{\frown}{,} 9$; Akören Plateau,, 18.04.2020, $1\stackrel{\frown}{,} 9$.

Comments: This Holomediterranean species was recorded on the *Alnus* sp., *Artemisia* sp., and *Geranium* sp. and from Çorum for the first time in the study. It is widespread and frequently distributed in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Yıldırım et al., 2011; Matocq et al., 2014; Fent & Dursun, 2019; Fent & Dursun, 2019; Zengin & Dursun, 2019; Altınok & Muştu, 2019).

Rhopalus subrufus (Gmelin, 1790)

Material examined: Kargı, Başköy, 26.06.2021,19, 13.

Comments: This Holomediterranean species was recorded from Çorum for the first time in the study. It is widespread and frequently distributed in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Uygun & Yiğit, 1982; Yıldırım et al., 2013; Beyaz & Tezcan, 2002; Fent & Dursun, 2019; Kıyak & Baş, 2021; Sarı & Yıldırım, 2021). It was found on the *Poa* sp.

Stictopleurus Stål, 1872

Stictopleurus abutilon (Rossi, 1790)

Material examined: Corum, Center, 18.08.2020, 19, 233; 06.06.2021, 19.

Comments: In the study, this Holomediterranean species was recorded for the first time from Çorum. It was found on the *Achillea* sp. and *Artemisia* sp. It is widespread and frequently distributed in Turkey (Puton, 1892; Horvath, 1901; Fahringer, 1922; Gadeau de Kerville, 1939; Tuatay et al., 1972; Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Yıldırım et al., 2011; Matocq et al.,, 2014; Fent & Dursun, 2019; Zengin & Dursun, 2019; Bolu, 2020; Bulak Korkmaz & Yıldırım, 2021).

Chorosoma Curtis, 1830

Chorosoma schillingii (Schilling, 1829)

Material examined: Çorum, Center, 06.06.2021, 299, 233; Dodurga, Berkköy, 18.09.2020, 299, 13; **Osmancık,** Akören Plateau, 18.04.2020, 299, 333; 25.05.2019, 399, 433; Ortaköy, Fındıklı, 26.07.2020, 19, 233; Sarpınkavak, 19.10.2020, 19 13.

Comments: This Holomediterranean species is widespread and frequently distributed in Turkey and it was recorded for the first time from Çorum province in this study (Horvath, 1894; 1905; Puton et Noualhier, 1895; Kiritshenko, 1918; Fahringer, 1922; Hoberlandt,1955; Pehlivan, 1981; Dursun, 2009; Yıldırım et al., 2013; Matocq et al., 2014; Fent & Dursun, 2019; Zengin & Dursun, 2019). This species has been found on the *Poa* sp.

Stenocephalidae Dallas, 1852

Dicranocephalus Hahn, 1826

Dicranocephalus agilis (Scopoli, 1763)

Material examined: Çorum, Center, 27.07.2021, 233; Osmancık, Akören Plateau, 18.04.2020, 1, 13; Kargı, Kargı Plateau, 19.06.2021, 1, 13.

Comments: This Holomediterranean species is widespread distributed and frequently found in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Fent & Dursun, 2019; Zengin & Dursun, 2019). The finding of the species is the first record for Corum province and it has been found on *Euphorbia* sp.

Dicranocephalus albipes (Fabricius, 1781)

Material examined: Bayat, 19.07.2020, 299; Kargı, Kargı Plateau, 19.06.2021, 299, 333; Kargı Tatil Köyü, 26.06.2021, 699, 433; Osmancık, Ardıç, 02.10.2021, 199; Öbektaş Plateau, 24.07.2021, 199; Sungurlu, Kertme, 10.07.2021, 499, 333.

Comments: The present record of this Holomediterranean species is the first time for the Heteroptera fauna of Çorum province. It has been found on *Euphorbia* sp. This species is widespread distributed and frequently found in Turkey (Pehlivan, 1981; Önder et al., 2006; Dursun, 2009; Küçükbasmacı & Kıyak, 2015; Fent & Dursun, 2019).

Saravdüzü Kuruçay D775 Vezirköprü Ceheli 32 Alanşeyhi 8 Pazarci 3 31 Ortalica 36 D031 Umac Kargın Gürmüşhacıköy Merzifon Yavu Yemicami Düwenci **E** 12 D180 D795 18 açat 28 Karakecili D180 Me 37 zü Yaydiğin 38 Uğ 30 ağ D785 Göynüc D795 alagöz Kızılırmak Dertli 13 10 Hüyük D785 Aydıncık

Figure 1. The area of study in Corum (from google earth).

CONCLUSION AND DISCUSSION

Delice

Coreoidea fauna of Corum province.

gracilicornis, study area. Ceraleptus species Coriomeris affinis, Centrocoris variegatus, In the present study, 1 species origin of Spathocera lobata from the family Coreidae Euro-Siberian, 20 species

and Brachycarenus tigrinus, Maccevethus errans caucasicus from the In this study, as a result of the identification Rhopalidae found at only in one locality of the material collected from 39 different in the research area too. Although of in around Corum province those species C. affinis, C. variegatus, B. between 2019 and 2021 revealed 3 tigrinus and M. errans caucasicus are species belonging to 2 genera of the widespread reported and frequently founded Alydidae family, 13 species belonging to in Turkey, the species were found rarely 10 genera of the Coreidae family, 10 in research area. Among the species of species belonging to 7 genera of the superfamily Coreoidea recorded in the Rhopalidae family and 2 species belon-surrounding of Corum; A. calcaratus, C. ging to 1 genera of the Stenocephalidae lateralis, C. marginatus, G. acuteangulatus, family. The total 28 species belonging to G. juniperi, P. laciniata, C. hyoscyami, R. 20 genera of the superfamily Coreoidea parumpunctatus, C. schillingii, D. albipes were reported in Corum province. Of and the invasive alien species L. occidentalis them, 23 species are new records for the are widespread distributed and frequently found in Turkey and also in our present

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Gököz

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Table 1. The plant relationships of Coreoidea species from Corum province

Family	Species	Origin	Host Plant
	Alydus calcaratus (Linnaeus, 1758)	Holomediterranean	Astragalus sp. (Fabaceae)
Alydidae Amyot & Serville, 1843 Coreidae Leach, 1815	Camptopus lateralis (Germar, 1817)	Holomediterranean	Chrysantemum sp. (Asteraceae) Rosa sp.,(Rosacae) Rumex sp. (Polygonaceae)
	Camptopus tragacanthae (Kolenati, 1845)	Irano-Turanian	Astragalus sp. (Fabaceae)
	Ceraleptus gracilicornis (Herrich-Schaeffer, 1835)	Holomediterranean	Trifolium sp. (Fabaceae)
	Coriomeris affinis (Herrich-Schaeffer, 1839)	Mediterranean	Pinus sp. (Pinaceae)
	Coriomeris denticulatus (Scopoli, 1763)	Euro-Siberian	Lamium sp. (Lamiaceae)
	Leptoglossus occidentalis Heidemann, 1910	Neactic region	Pinus sp. (Pinaceae)
	Centrocoris spiniger (Fabricius, 1781)	Holomediterranean	Cirsium sp. (Asteraceae)
	Centrocoris variegatus Kolenati, 1845	Holomediterranean	Cirsium sp. (Asteraceae)
	Coreus marginatus (Linnaeus, 1758)	Holomediterranean	Rumex sp. (Polygonaceae)
	Enoplops disciger (Kolenati, 1845)	Pontomediterranean	Echium sp. (Boraginaceae)
	Spathocera lobata (Herrich-Schaffer, 1840)	Holomediterranean	Rumex sp. (Polygonaceae)
	Syromastus rhombeus (Linnaeus, 1767)	Holomediterranean	Poa sp. (Poaceae)
	Gonocerus acuteangulatus (Goeze, 1778)	Holomediterranean	Juniperus sp. (Cupressaceae)
	Gonocerus juniperi Herrich-Schaeffer, 1839	Holomediterranean	Juniperus sp. (Cupressaceae)
	Phyllomorpha laciniata (Villers, 1789)	Holomediterranean	Acantholimon sp. (Plumbaginaceae) Astragalus sp. (Fabaceae)
	Brachycarenus languidus (Horváth, 1891)	Irano-Turanian	Chenopodium sp. (Chenopodiaceae)
	Brachycarenus tigrinus (Schilling, 1829)	Holomediterranean	Chenopodium sp. (Chenopodiaceae)
	Corizus hyoscyami (Linnaeus, 1758)	Holomediterranean	Hypericum sp. (Hypericaceae) Onosma sp. (Boraginaceae)
Rhopalidae Amyot&Serville, 1843	Liorhyssus hyalinus (Fabricius, 1794)	Holomediterranean	Poa sp. (Poaceae)
	Maccevethus corcicus corsicus Signoret, 1862	Pontomediterranean	Poa sp. (Poaceae)
	Maccevethus errans caucasicus (Kolenati, 1845)	Pontomediterranean	Erodium sp. (Geraniaceae)
	Rhopalus parumpunctatus Schilling, 1829	Holomediterranean	Alnus sp., (Betulaceae) Artemisia sp., (Asteraceae) Geranium sp. (Geraniaceae)
	Rhopalus subrufus (Gmelin, 1790)	Holomediterranean	Poa sp. (Poaceae)
	Stictopleurus abutilon (Rossi, 1790)	Holomediterranean	Achillea sp., Artemisia sp. (Asteraceae)
	Chorosoma schillingii (Schilling, 1829)	Holomediterranean	Poa sp. (Poaceae)
Stenocephalidae	Dicranocephalus agilis (Scopoli, 1763)	Holomediterranean	Euphorbia sp. (Euphorbiaceae)
Dallas, 1852	Dicranocephalus albipes (Fabricius, 1781	Holomediterranean	Euphorbia sp. (Euphorbiaceae)

Holomediterranean, 2 species origin of Nearctic region (cosmopolitan) have been Irano-Turanian, 1 species origin of Medi- recorded from Çorum. Holomediterranean, 3 species origin of Ponto- terranean species are a very common and mediterranean, and 1 species origin of abundant in the surroundings of Çorum.

As a result of our observations in the field, a total of 21 taxon at genus level from 15 plant families were identified in relation to Coreoidea species. When take into account the preferences of Coreoidea Bulak Korkmaz, Y., Yıldırım, E., 2021, Contribution species in these families, 5 species were associated with Asteraceae and Poaceae. 4 species with Fabaceae, 3 species with Polygonaceae, 2 species with Boraginaceae, Chenopodiaceae, Cupressaceae, Euphorbiaceae, Geraniaceae and Pinaceae, 1 species with Betulaceae, Hypericaceae, Plumbaginaceae Çerçi, B., Özgen, İ., 2021, Contribution to the and Rosacae (Table.1).

Many microclimate areas is very important for faunal component. The geographical location of Corum is beetwen central Antolia and central Black sea region. Cerçi, B., Özgen, İ., Dioli, P., 2016, Additional There are also microclimate areas Corum. Because of its location, Corum is a dispersal corridor for different animal. With the present additional records were contributed to the distribution of the families Alydidae, Coreidae, Rhopalidae Stenocephalidae, in Turkey the determination of biodiversity of this fami- Dolling, W. R., 2006, Coreidae Leach, 1815. lies in the surroundings of Corum.

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AUTHOR GUIDELINES Aims & Scope

The *Journal of the Heteroptera of Turkey* is a biannual peer reviewed international journal that publishes original articles, review articles, and short communication on all aspects of Heteroptera.

The *Journal of the Heteroptera of Turkey* publishes qualified research articles on the systematics, taxonomy, faunistical and ecology of heteroptera suborder. The topic of the research may include a wide range of heteropteran fields. Detailed studies on systematics, morphology, ecology, and phenology of heteroptera, and the biological, ecological, and faunistic formation of heteroptera taxons.

In this Journal full-papers and short communications containing original researches on any aspect of heteropteran in palaearctic region and Turkey will be considered as publication.

The Journal of the Heteroptera of Turkey welcomes review articles in the field of heteropteran.

The *Journal of the Heteroptera of Turkey* also published short notes on heteropteran topics. Information of the heteropteran specialists and book reviews will also be published.

We would like to make an open invitation to all potential contributors. We have a fast publishing process to process and evaluate.

Taxonomic revisions and descriptions of individual species will be accepted especially if additional information is included on habitat preferences, behavior, phenology etc. Descriptions of single specimens are discouraged.

For submitted article there are restrictions on the subject, author, geographic area, and so on of any submission (palaearctic only). For our journal mission all fields of heteroptera studies are suitable.

All papers being peer-reviewed by two referees, and under rapid publication process.

Preparation of Manuscript

All manuscripts should be written in the Turkish or English languages to be published only in the Journal of the Heteroptera and should be prepared with Microsoft Word.

Manuscripts should be written on A4 (21 cm x 29.7 cm) paper with margins of at least 2 cm in width.

All pages should be numbered consecutively. Manuscripts should be organized in the following order: Title, abstract, brief introduction, materials and methods, results, discussion, acknowledgments, references, tables and figure legends.

Parts of the Manuscript should be:

Arrange manuscripts in this order: title; name(s), address(es) and e-mail address(es) of the corresponding author(s) who will receive and approve the page proofs (research articles only); keywords; text; acknowledgments; references; tables and figure legends.

Title: The title of the manuscript should be informative and clear, not exceed 15-20 words. Just under the title full name(s) of author(s); (surname(s) in capital letters; full address(es); e-mail address(es); if available, ORCID numbers for all authors, Corresponding Author contact information should be give (each on a separate line).

Abstract: The abstract should not exceed 250-300 words (maximum), should be one paragraph.

Keywords: For subject indexing, up to 6 topical keywords in English are required (for Turkish articles).

Text: Introduction, Materials and Methods, Results, Conclusion and Discussion, Acknowledgments, References, Figure and table legends.

Use italics for Scientific names of genera, species, and subspecific taxa.

Do not use italics for abbreviations such as "spp.", "sp.", "sp.", "var.", "gen.nov.", "sp.n.", "ssp.nov.", "stat.n.", "comb.n.", "s.l.", "s. str.", "et al.", and names of taxa of rank higher than genus.

For faunistic research follow this order: Taxon name, Material examined, Habitat, Host plant(s), Distribution. Example:

Miridae Hahn, 1831

Deraeocoris rutilus (Herrich-Schaeffer, 1838)

Habitat: The specimens belonging to *D. rutilus* (H.,-S., 1838) were found on *Carduus pycnocephalus* subsp. *albidus* (Bieb) Kazmi.

Materials examined: 1 male, 24.6.1996 (Loc. 1), 1 female, 24.6.1996 (Loc.6).

Distribution in Turkey: The Aegean, the Marmara, and the Anatolia regions (18,10,8,13,29). Distribution in the world: Israel, Sardinia, Syria, Cyprus, Poland, the Balkans, Russia, and Turkey (18,25).

References: References should be prepared according to "The Guidelines to Authors".

The complete reference list should appear alphabetically by name at the end of the paper. A sample of the most common entries in reference lists appears below. Please note that a DOI should be provided for all references where available.

References must be cited in the text as (Dursun, 2013), Fent & Dursun (2005) or Fent et al. (1997), or in a parenthesis (Dursun, 2013; Fent & Dursun, 2005; or Fent et al. ,1997).

Journal article: Abbreviate names of periodicals basically according to the World List of Scientific Periodicals, 4th Edition, Butterworths, London, 1964–1965. (If you are not certain about the correct abbreviation, give the journal's name in full).

Fent, M., Kment, P., Elipek-Çamur, B., Kırgız, T., 2011, Annotated catalogue of Enicocephalomorpha, Dipsocoromorpha, Nepomorpha, Gerromorpha and Leptopodomorpha (Hemiptera: Heteroptera) of Turkey with new records, *Zootaxa*, 2856:1-84.

Books: Alexi Popov, A., Grozeva, S., Simov, N., Tasheva, E., 2013, *Advances in Hemipterology*, PenSoft Publishers Ltd., 377 pp., Sofia, Bulgaria.

Article/Chapter in Book: Kerzhner, I. M., Jaczewski, T. L.,1964, *Order Hemiptera (Heteroptera)* 851–1118pp. In: Keys to the insects of the European USSR 1. (Ed. G. Y. Bei-Bienko). Nauka, Moskva & Leningrad [in Russian; English translation, Israel Program for Scientific Translations, Jerusalem, 1967]. 1214 pp.

No Author Given: (USDA) U.S. Department of Agriculture. 2001. Title. USDA, Beltsville, MD. (IRRI) International Rice Research Institute. 2001. Title. IRRI, City, State or Country.

Proceedings: Šeat, J., Kaur, H., Gallé, R., Torma, A. 2018, The role of road verges as secoundary linear habitats for Forest steppe Heteroptera, 8th European Hemiptera Congress, 24-29 Jun 2018, Zawiercie, Poland. Book of Abstracts, 61 p.

Theses/Dissertations: James, H., 2001, Thesis or dissertation title. M.S. thesis or Ph.D. dissertation, University of Pennsylvania, Philadelphia.

Özsaraç, Ö., 2004, Çiçekdağı (Kırşehir) Heteropterleri, Basılmamış Doktora Tezi, Gazi Üniversitesi Fen Bilimleri Enstitüsü, Ankara, 225 s.

Online Citations/ Websites: Rabitsch, W., 2005, Spezialpraktikum aquatische und semiaquatische Heteroptera; (Web page: http://homepage.univie.ac.at/wolfgang.rabitsch).(Date accessed: May 2010).

Using the DOI (Digital Object Identifier) Number: Nestel D., Papadopoulos N. T. & Miranda Chueca M. A. (2008). Current advances in the study of the ecology of fruit flies from Europe, Africa and the Middle East. *Journal of Applied Entomology*, DOI: 10.1111/j.1439-0418.2008.01378.x

Please note on the illustrations, figure, table, and photographs legends: Illustrations should be arranged into blocks or plates by the author(s). Figures should be provided electronically in either JPG or TIFF format. JPG images should be the highest resolution possible. TIFF images should be at 300 dpi resolution.

Morphological illustrations (if not schematic) and **photographs**/ electron microscope micrographs should include scale bars. Photographs and electron microscope micrographs must be in JPEG file format (300 dpi).

Images pasted into Word become low-resolution and cannot be used in print.

Photographs should be high-contrast, black and white or color. Lettering should be typed and legible. All papers should be accompanied by information on the credited photographer or copyright holder. If the photographer or copyright holder is not an author on the paper, then permission must be granted by the copyright holder.

Tables should be numbered consecutively and include headings and explanations. References in the text to illustrations (schematic, photographs) and tables into parenthesis: e.g.(Fig.1) (Figs.1-4) (Table 1.) (Table 1., Figs.1-4). Morphological illustrations should be provided with scale bars.

Taxonomic papers in JHT must follow the requirements below: Follow all requirements of the current International Code of Zoological Nomenclature (4th edition 1999), and be followed the recommendations of the Code.

A holotype should always be designated for each newly described species-level taxon and at least holotypes should be deposited in public collections that provide long-term care and access for study (note that such deposition is mandatory for neotypes). For this reason, two particular recommendations (73A and 16C) should be observed in JHT.

In the Abstract must be listed new combinations, new status, new taxa, new synonyms, etc. in. The list of synonymized names must indicate their disposition. For newly should be described taxa included for all newly synonymized or combined names. Use "sp. n.", "gen. n." etc.. **Important note:** Descriptions based on single specimens are discouraged.

The standard order of sections for description a species is: "Diagnosis", "Description", "Material", "Type locality", "Etymology", "Distribution", "Biology", and other comments if appropriate. Author(s) of species name must be provided when the scientific name of any animal species is first mentioned. (The year of publication is not compulsory. If you give it, then provide a full reference of this in the reference list.)

It is the *author's responsibility* to know the group, both material and literature, well enough (preferably on a worldwide basis) to be able to ensure that all relevant taxa were taken into account and that any new taxa proposed have not already been described from elsewhere.

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